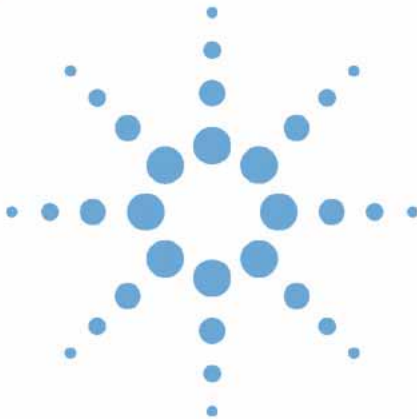




Agilent 1200 Series Micro Collector/Spotter



User Manual



Agilent Technologies

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CAUTION

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WARNING

A **WARNING** notice denotes a hazard. It calls attention to an operating procedure, practice, or the like that, if not correctly performed or adhered to, could result in personal injury or death. Do not proceed beyond a **WARNING** notice until the indicated conditions are fully understood and met.

In This Guide...

This manual contains user information about the Agilent 1200 Series micro collector/spotter G1364D. It is intended for users that will operate the Agilent 1200 Series micro collector/spotter either for micro fraction collection or for MALDI Spotting. For information about installation, advanced troubleshooting, repair and a complete list of internal parts please refer to the *Agilent 1200 Series Fraction Collector Service Manual G1364-90110*.

1 Installing the Micro Collector/Spotter

This chapter provides information about the installation of the Agilent 1200 Series micro collector/spotter.

2 Configuration and Operation of the Micro Collector/Spotter

This chapter describes the configuration, method setup and operation of the micro collector/spotter.

3 Control Module Screens of the Fraction Collector

The reference to the screens of the Agilent 1200 Series control module is provided in the chapter.

4 Troubleshooting and Test Functions

This chapter describes the modules built-in troubleshooting, test and maintenance functions.

5 Maintenance and Simple Repairs

This chapter contains instructions on maintenance and simple repair procedures.

6 Parts and Materials

This chapter contains selected illustrations and lists for identification of parts and materials that are required for maintenance, or exchanged in simple repair procedures.

7 Specifications

This chapter contains performance specifications of the micro collector/spotter.

A Safety Information

This appendix provides a safety summary.

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Installing the Micro Collector/Spotter

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Site Requirements

A suitable site environment is important to ensure optimum performance of the micro collector/spotter.

Power Consideration

The micro collector/spotter power supply has wide-ranging capability (see [Table 1](#) on page 12). Consequently there is no voltage selector in the rear of the micro collector/spotter. There are also no externally accessible fuses, as automatic electronic fuses are integrated in the power supply.

The thermostatted micro collector/spotter comprises two modules, the micro collector/spotter (G1364D) and the thermostat (G1330B). Both modules have a separate power supply and a power plug for the line connections. The two modules are connected by a control cable and both are turned on by the micro collector/spotter module. The thermostat power supply has two externally accessible fuses.

WARNING

To disconnect the micro collector/spotter from line power, unplug the power cord. The power supply still uses some power, even if the power switch on the front panel is turned off.

WARNING

To disconnect the thermostatted micro collector/spotter from line power, unplug the power cord from the micro collector/spotter and the thermostat. The power supplies still use some power, even if the power switch on the front panel is turned off. Please make sure that it is always possible to access the power plug.

WARNING

Shock hazard or damage of your instrumentation can result if the devices are connected to a line voltage higher than specified.

Power Cords

Your micro collector/spotter is delivered with a power cord which matches the wall socket of your particular country or region. The plug on the power cord which connects to the rear of the instrument is identical for all types of power cord.

WARNING

Never operate your instrumentation from a power outlet that has no ground connection. Never use a power cord other than the power cord designed for your region.

WARNING

Never use cables other than the ones supplied by Agilent Technologies to ensure proper functionality and compliance with safety or EMC regulations.

Bench Space

The micro collector/spotter dimensions and weight (see [Table 1](#) on page 12) allow the instrument to be placed on almost any laboratory bench. The instrument requires an additional 2.5 cm (1.0 inch) of space on either side, and approximately 8 cm (3.1 inches) at the rear for the circulation of air, and room for electrical connections. Ensure the micro collector/spotter is installed in a horizontal position.

The thermostatted micro collector/spotter dimensions and weight (see [Table 2](#) on page 13) allow the instrument to be placed on almost any laboratory bench. The instrument requires an additional 25 cm (10 inches) of space on either side for the circulation of air, and approximately 8 cm (3.1 inches) at the rear for electrical connections. Ensure the micro collector/spotter is installed in a horizontal position.

If a complete Agilent 1200 Series LC system is to be installed on the bench, make sure that the bench is designed to carry the weight of all the modules. For a complete system including the thermostatted micro collector/spotter it is recommended to position the modules in two stacks, see [“Recommend Stack Configuration for micro collector/spotter in the capillary or nanoflow system”](#) on page 18. Make sure that in this configuration there is 25 cm (10 inches) space on either side of the thermostatted micro collector/spotter for the circulation of air.

Environment

Your micro collector/spotter will work within specifications at ambient temperatures and relative humidity as described in [Table 1](#) and [Table 2](#) on page 13.

CAUTION

Do not store, ship or use your micro collector/spotter under conditions where temperature fluctuations may cause condensation within the micro collector/spotter. Condensation will damage the system electronics. If your micro collector/spotter was shipped in cold weather, leave it in its box, and allow it to warm up slowly to room temperature to avoid condensation.

Table 1 Physical Specifications - micro collector/spotter G1364D

Type	Specification	Comments
Weight	13.5 kg (29.8 lbs)	
Dimensions (height × width × depth)	200 × 345 × 440 mm (8 × 13.5 × 17 inches)	
Line voltage	100 – 240 VAC, ±10 %	Wide-ranging capability
Line frequency	50 or 60 Hz, ±5 %	
Power consumption (apparent power)	200 VA	Maximum
Power consumption (active power)	180 W	Maximum
Ambient operating temperature	4 – 55 °C (41 – 131 °F)	
Ambient non-operating temperature	-40 – 70 °C (-4 – 158 °F)	
Humidity	< 95 %, at 25 – 40 °C (77 – 104 °F)	Non-condensing
Operating Altitude	Up to 2000 m (6500 ft)	
Non-operating altitude	Up to 4600 m (14950 ft)	For storing the micro collector/spotter
Safety standards: IEC, CSA, UL	Installation Category II, Pollution Degree 2. For indoor use only.	

Table 2 Physical Specifications - Thermostat (G1330B)

Type	Specification	Comments
Weight	18.5 kg (40.7 lbs)	
Dimensions (height × width × depth)	140 × 345 × 435 mm (5.5 × 13.5 × 17 inches)	
Line voltage	100 – 120 or 220 – 240 VAC, ± 10 %	Wide-ranging capability
Line frequency	50 or 60 Hz, ± 5 %	
Power consumption (active power)	210 W	Maximum
Power consumption (apparent power)	260 VA	Maximum
Ambient operating temperature	4 – 40 °C (41 – 131 °F)	
Ambient non-operating temperature	-40 – 70 °C (-4 – 158 °F)	
Humidity	< 95 %, at 25 – 40 °C (77 – 104 °F)	Non-condensing
Operating Altitude	Up to 2000 m (6500 ft)	
Non-operating altitude	Up to 4600 m (14950 ft)	For storing the micro collector/spotter
Safety standards: IEC, CSA, UL, EN	Installation Category II, Pollution Degree 2. For indoor use only.	

Unpacking the Fraction Collector

CAUTION

If you need to ship the micro collector/spotter at a later date, always use the shipping protection foam parts (see [“Transporting the Micro Collector Spotter”](#) on page 32).

Damaged Packaging

Upon receipt of your micro collector/spotter, inspect the shipping containers for any signs of damage. If the containers or cushioning material are damaged, keep them until the contents have been checked for completeness and the micro collector/spotter has been mechanically and electrically checked. If the shipping container or cushioning material is damaged, notify the carrier and keep the shipping material for the carrier’s inspection.

CAUTION

If there are signs of damage to the micro collector/spotter, please do not attempt to install the micro collector/spotter.

Delivery Checklist

Ensure all parts and materials have been delivered with the micro collector/spotter. For this compare the shipment content with the checklist included in each instrument box. Please report missing or damaged parts to your local Agilent Technologies sales and service office.

The Agilent 1200 Series micro collector/spotter is available as:

- *G1364D* micro collector/spotter, designed for flow rates below 100 µl and for use with well plates and Eppendorf tubes.
- *G1364D Thermostatted* option of the micro collector/spotter. This option can be setup by additionally ordering and installing a G1330B fraction collector thermostat

Table 3 Delivery Checklist for the G1364D micro collector/spotter

Description	Quantity
micro collector/spotter unit	1
Power cord, local (Matching the sockets in your country or region)	1
Accessory kit (see below)	1

Table 4 Delivery Checklist for the G1330B Thermostat Module (optional)

Description	Quantity
Thermostat Module	1
Power cord, local (Matching the sockets in your country or region)	1
Accessory kit thermostat module (see below)	1

Accessory Kits

Each shipment contents an Accessory kit with the necessary tools to install the system and to have an operating system.

- The Accessory kit (G1364-68715) shown in [Table 5](#) is shipped with the micro collector/spotter (G1364D).

Table 5 Micro collector/spotter Accessory Kit Contents G1364-68715

Description	Quantity	Part Number
Hex key 2.0 mm	1	8710-2476
MALDI Spotting Adapter	1	G1364-83205
Flap Septum	1	G1364-27107
Waste Tubing 1.4 mm ID	1	G1364-86711
Waste tubing (1.2 m)*	1	5062-2463
CAN cable, 1 m	1	5181-1519

1 Installing the Micro Collector/Spotter

Unpacking the Fraction Collector

Table 5 Micro collector/spotter Accessory Kit Contents G1364-68715

Description	Quantity	Part Number
Sticking clamp for corrugated waste tubing (large) [†]	3	5065-9976
Sticking clamp for waste tubing (small) [‡]	3	5065-9976
ESD wrist strap	1	9300-1408
micro collector/spotter Capillary Assembly (25 µm ID)	1	G1364-87304
micro collector/spotter Capillary Assembly (100 µm ID)	1	G1364-87306
ZDV Union, stainless steel	1	5022-2184
Strip with 10 MALDI Spotting Tips	1	G1364-81701
Plastic Spacers	20	not orderable
Plug for waste container	1	G1364-26105

* Reorder gives 5 m

† Reorder gives 10 clamps for corrugated waste tubing (large) and 10 clamps for waste tubing (small)

‡ Reorder gives 10 clamps for corrugated waste tubing (large) and 10 clamps for waste tubing (small)

- The Accessory kit (G1330-68705) shown in [Table 6](#) is shipped with the (G1330B) thermostat module if the thermostat module was ordered.

Table 6 (Optional) Thermostat Module Accessory Kit Contents G1330-68705

Description	Quantity	Part Number
Waste tubing (1.2 m) [*]	1	5062-2463
Cable thermostat module to micro collector/spotter	1	G1330-81600

* Reorder gives 5 m

Optimizing the Stack Configuration

If your Agilent 1200 Series micro collector/spotter is part of a system, you can ensure optimum performance and minimum delay volume by installing the following configuration.

- [Figure 1](#) on page 18 shows the configuration recommended for the micro collector/spotter within an Agilent 1200 Series capillary or nanoflow LC system

1 Installing the Micro Collector/Spotter

Optimizing the Stack Configuration

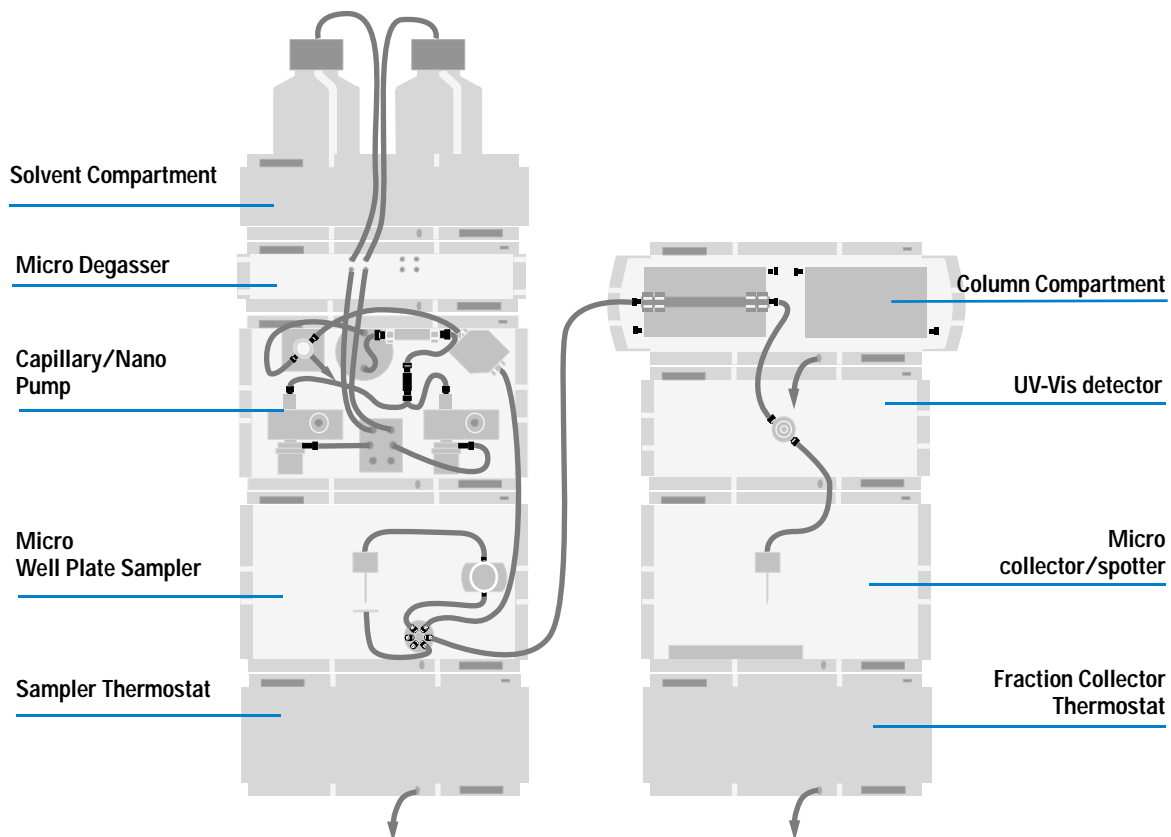


Figure 1 Recommend Stack Configuration for micro collector/spotter in the capillary or nanoflow system

NOTE

The micro collector/spotter should never be positioned on top of a module that generates heat, which can lead to an unwanted evaporation of fractions in the micro collector/spotter (e.g. Agilent 1200 Series thermostatted column compartment G1316A/B or Agilent 1200 Series diode-array detectors G1315B/C).

Installing the Micro Collector/Spotter

Preparation	Locate bench space Provide power connections Unpack the micro collector/spotter
Parts required	Fraction Collector Power cord Chemstation and/or Control Module G1323B

WARNING

When opening capillary or tube fittings solvents may leak out. Please observe appropriate safety procedures (for example, goggles, safety gloves and protective clothing) as described in the material handling and safety data sheet supplied by the solvent vendor, especially when toxic or hazardous solvents are used.

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation. Do not attempt to insert or remove a vial or a plate when the needle is positioned.

- 1 Install the LAN interface board in the micro collector/spotter (if required).
- 2 Remove the adhesive tape which covers the side and front doors.
- 3 Open the front door and remove the left side door.
- 4 Remove the transport protection foam.
- 5 Install the corrugated waste tube in the plastic port at the front bottom center of the instrument. Slide the waste tubing coming from the internal tray (if present) through the plastic port and the corrugated waste tube (see [Figure 2](#) on page 20). Route the corrugated waste tubing into a waste container.

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Installing the Micro Collector/Spotter

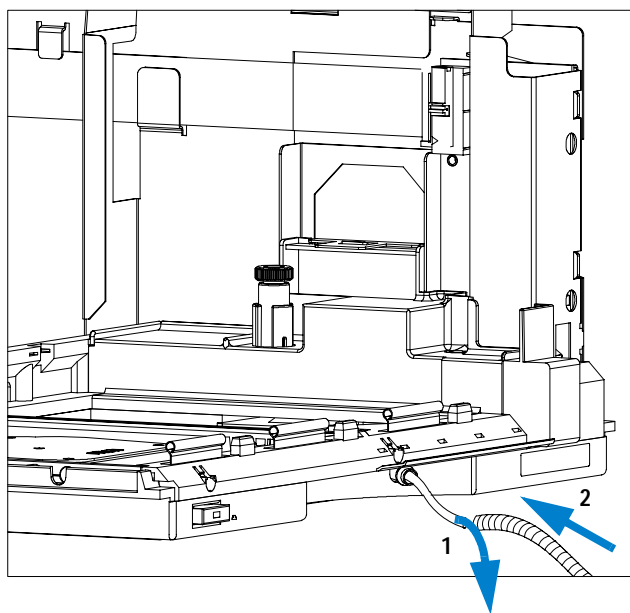


Figure 2 Installing the Corrugated Waste Tubing in the Plastic Port

- 6 Re-install the left side door (take care of the magnet at the back). Ensure the side door is correctly installed (its presence is sensed by a hall sensor, a missing side door will result in a NOT-READY state of the instrument).
- 7 Place the micro collector/spotter in the stack or on the bench in all horizontal position.
- 8 Ensure the power switch at the front of the micro collector/spotter is OFF.
- 9 Connect the power cable to the power connector at the rear of the micro collector/spotter.
- 10 Connect the CAN cable to the other Agilent 1200 Series LC modules.
- 11 If an Agilent ChemStation is the controller, it must be connected to the LAN interface (should be installed to the detector)
- 12 Connect the APG remote cable (optional) for non Agilent 1200 Series instruments.
- 13 Install the tray that has been delivered for your micro collector/spotter.

NOTE

The micro collector/spotter (G1364D) is shipped with 4 x well plate full tray. All other trays have to be ordered separately.

- 14 Turn ON power by pushing the button at the lower left hand side of the micro collector/spotter.
- 15 The exhaust fan will turn ON and remove potential solvent vapor from the inside of the instrument. After 2 minutes close the front door. Then the micro collector/spotter will start the hardware initialization process. At the end of this process the status LED should be green.

Installing the Micro Collector/Spotter

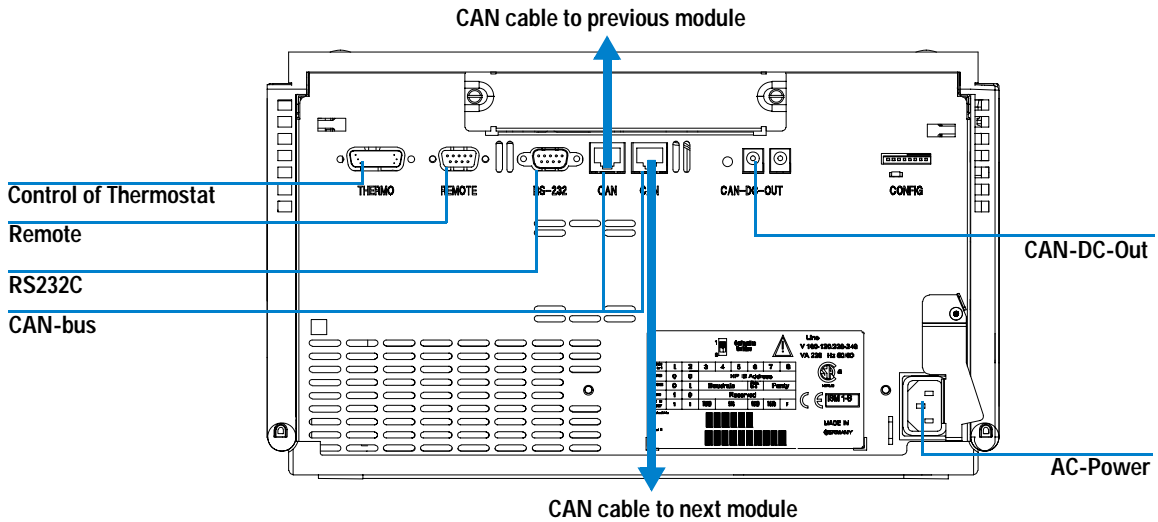


Figure 3 Cable Connections

NOTE

The micro collector spotter is turned ON when the line power switch is pressed and the green indicator lamp is illuminated. The detector is turned OFF when the line power switch is protruding and the green light is OFF.

WARNING

To disconnect the micro collector/spotter from the line, unplug the power cord. The power will supply still uses some power, even switch at the front panel is turned OFF.

Installing a Thermostatted Fraction Collector

Preparation	Locate bench space Provide power connections Unpack the micro collector/spotter and the thermostat
Parts required	Fraction Collector and thermostat Power cords ChemStation and/or Control Module G1323B

- 1 Place the thermostat on the bench.
- 2 Remove the front cover and route the condensation drain tube to a waste container.

WARNING

Make sure the condensation drain tube runs down into a waste container without any (upwards) bends or curves. Free and unrestricted flow of the condensation into a waste container must be guaranteed. Make sure that the condensation drain tube is always above the liquid level in the container. If the tube is located in liquid the condensed water cannot flow out of the tube and the outlet is blocked. Any further condensation will then remain in the instrument. This may damage the instruments electronics.

CAUTION

The micro collector/spotter thermostat requires 25 cm (10 inch) space on each for sufficient air circulation.

- 3 Install the LAN interface board in the micro collector/spotter (if required).
- 4 Remove the adhesive tape which covers the side and front doors.
- 5 Open the front door and remove the left side door.
- 6 Remove the transport protection foam.
- 7 Install the corrugated waste tube in the plastic port at the front bottom center of the micro collector/spotter and route down into a waste container. Slide the waste tubing coming from the internal tray (if present)

1 Installing the Micro Collector/Spotter

Installing a Thermostatted Fraction Collector

through the plastic port and the corrugated waste tube (see [Figure 2](#) on page 20). Route the corrugated waste tubing into a waste container.

- 8 Re-install the left side door (take care of the magnet at the back). Ensure the side door is correctly installed (its presence is sensed by a hall sensor, a missing side door will result in a NOT-READY state of the instrument).
- 9 Place the micro collector/spotter on top of the thermostat. Make sure that the micro collector/spotter is correctly engaged in the thermostat locks.
- 10 Remove the plastic cover from the tray base, place the air channel adapter (1) into the micro collector/spotter tray base. Make sure the adapter is fully pressed down. This assures that the cold airstream from the thermostat is correctly guided to the tray area of the micro collector/spotter. Place the plug channel (2) on top of the air channel adapter. Both devices must be installed correctly, to assure proper operation of the instrument.

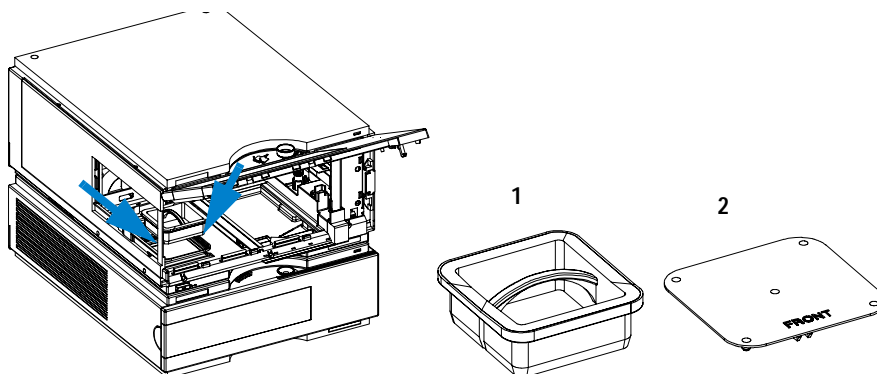


Figure 4 Installation of Thermostat and Fraction Collector

- 11 The Agilent 1200 Series micro collector/spotter is delivered with a pre-installed tray compartment divider. This divider should only be used with the thermostatted micro collector/spotter, if a (half or std.) tray is installed into the left and center positions of the instrument. If any tray is installed to the right side of the instrument, remove the tray compartment divider. The tray compartment divider optimizes the cooling performance of the instrument, if only the left and center position of the micro collector/spotter are in use.

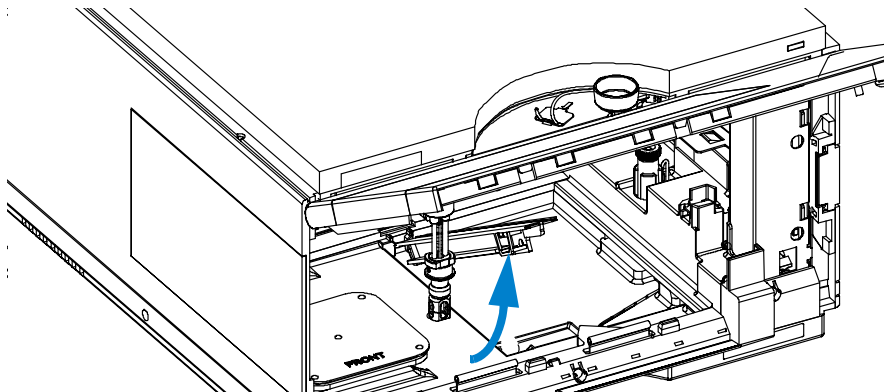


Figure 5 Removing the Tray Compartment Divider

- 12 Install the tray you have ordered for your micro collector/spotter.
- 13 Ensure the power switch on the front of the micro collector/spotter is OFF and the power cables are disconnected.
- 14 Connect the cable between the micro collector/spotter and the thermostat, see [Figure 6](#) on page 26.

WARNING

Do not disconnect or reconnect the micro collector/spotter to thermostat cable when the power cords are connected to either of the two modules. This will damage the electronics of the modules.

- 15 Connect the power cables to the power connectors.
- 16 Connect the CAN cable to other Agilent 1200 Series LC modules.
- 17 If an Agilent ChemStation is the controller, it must be connected to the LAN interface.
- 18 Connect the APG remote cable (optional) for non Agilent 1200 Series instruments.
- 19 Turn ON power by pushing the button at the lower left hand side of the micro collector/spotter.
- 20 The exhaust fan will turn ON and remove potential solvent vapor from the inside of the instrument. After 2 minutes close the front door. Then the

1 Installing the Micro Collector/Spotter

Installing a Thermostatted Fraction Collector

micro collector/spotter will start the hardware initialization process. At the end of this process the status LED should be green.

- 21 The micro collector/spotter is turned ON when the line power switch is pressed and the green indicator lamp is illuminated. The detector is turned OFF when the line power switch is protruding and the green light is OFF.

WARNING

To disconnect the micro collector/spotter from the line, unplug the power cord. The power supply still uses some power, even if the power switch at the front panel is turned OFF.

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation. Do not attempt to insert or remove a vial or a plate when the needle is positioned.

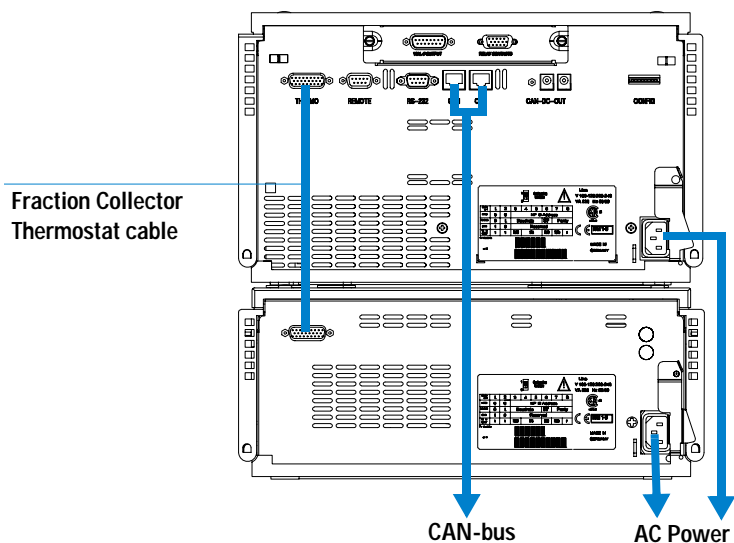


Figure 6 Connections at the Rear of the Thermostatted Fraction Collector

Micro Collector/Spotter Trays

Installing the Fraction Collector Trays

- 1 Press the front door latch-holding button located at the front of the right-side cover.
- 2 Lift the front door.
- 3 Adjust the top plate of the test tube tray for the correct tube height if required.
- 4 Load the micro collector/spotter tray with micro collector/spotter well-plates, test tubes or vials as required.
- 5 Slide the micro collector/spotter tray into the micro collector/spotter so that the rear of the tray is seated firmly against the rear of the tray area.

NOTE

Installed trays are automatically detected and identified.

- 6 Press the front of the micro collector/spotter tray down to secure the tray in the micro collector/spotter.
- 7 Close the front door.

NOTE

If the tray pops out of position the air channel adapter is not correctly inserted.

NOTE

Before starting a run, the instrument has to be correctly configured in the user interface.

Numbering of vial, well-plate and spot positions

With the 4 plates full tray

Plate in the left front position: P1

Plate in the left back position: P2

Plate in the right front position: P3

Plate in the right back position: P4

Vessel: A1; A2;... B1; B2;

With the 2 plates / 10 x 2ml vials or 10 funnels std. trays

Plate in the front position: P1

Plate in the back position: P2

Vessel: A1; A2;... B1; B2;...

Vials / funnels: 1 - 10

With the 100 vials std. tray

Vial: 1 - 100

With the half-trays

Left-hand 40-position tray: 1 - 40

Center 40-position tray: 101-140

Right-hand 40-position tray: 201 - 240

or

Left-hand 15-position tray: 1 - 15

Center 15-position tray: 101-115

Right-hand 15-position tray: 201 - 215

WARNING

Do not use closing mats. The micro collector/spotter has no needle, therefore the capillary will be bend or clogged after several injections.

Configure Well-plate Types

If the plate you are using is not found on the “[List of Recommended Plates](#)” on page 130 you may configure a custom plate. Measure the exact dimensions of the plate as marked below and enter the values in the plate configuration table of the ChemStation or the Control Module.

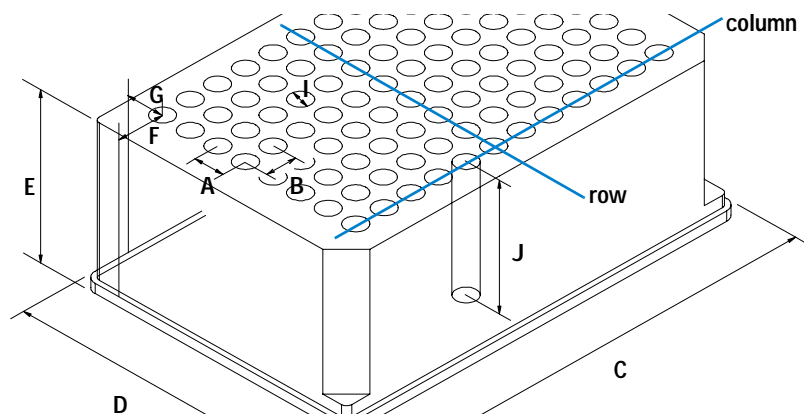


Figure 7 Well-plate Dimensions (straight)

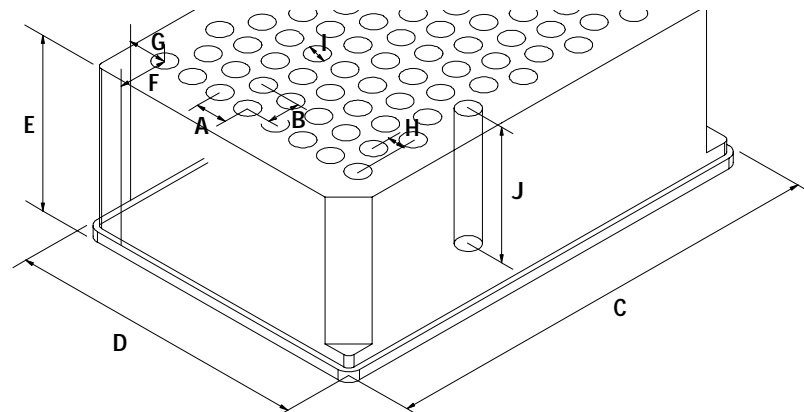


Figure 8 Well-plate Dimensions (staggered)

1 Installing the Micro Collector/Spotter

Configure Well-plate Types

Table 7 Configuring Well-plate Types

Location	Description	Definition	Limits
	Rows	Number of rows on the plate	up to 16
	Columns	Number of columns on the plate	up to 24
	Volume	Volume (in μl) of a sample vessel	
A	Row distance	Distance (in mm) between the center of two rows	
B	Column distance	Distance (in mm) between the center of two columns	
C	Plate length	X size (in mm) at the bottom of the plate	127.75+/- 0.25 mm (SBS Standard)
D	Plate width	Y size (in mm) at the bottom of the plate	85.50+/-0.25 mm (SBS Standard)
E	Plate height	Size (in mm) from the bottom to the top of the plate. If you use well plates with inlets, caps and septa, you have to specify the distance from the bottom to the top of the caps.	up to 47 mm
F	Column offset	Distance (in mm) from the left edge (bottom) to the center of the first hole (A1)	
G	Row offset	Distance (in mm) from the back edge (bottom) to the center of the first hole (A1)	
H	Column shift	Offset (in mm) to Y when the rows are not straight but staggered	
J	Well depth	Distance (in mm) from the top of the plate to the bottom of the well. If you use well plates with inlets, caps and septa, you have to specify the distance from the top of the septa to the bottom of the inlets.	up to 45 mm

Table 7 Configuring Well-plate Types (continued)

Location	Description	Definition	Limits
	Well X size	Size of the well in x direction (Plate length) If you use well plates with inlets, caps and septa, you have to specify the x size of the septa.	min. 3.7 mm min. 3.0 mm with position accuracy alignment (micro scale)
	Well Y size	Size of the well in y direction (Plate width). If you use well plates with inlets, caps and septa, you have to specify the y size of the septa.	down to 3.7 mm min. 3.0 mm with position accuracy alignment (micro scale)
	Bottom size	For round wells, the relative of the top and bottom of the well	1.0: cylindrical well 0.0: conical well
	Square	Click in the field to specify whether the well is rectangular or round	Yes: rectangular No: round /oval
	Is well plate	Click in this field to specify if this is a well plate or not. Relevant for continuous flow operation.	Yes: well plate or MALDI Target No: Vial Tray or Eppendorf tray

NOTE

The distances need to be measured with high precision. It is recommended to use a caliper. If possible contact the vendor of non-predefined well plates to obtain these dimensions.

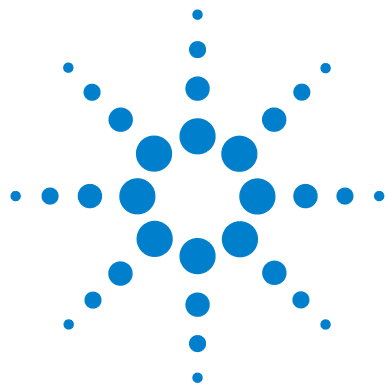
Transporting the Micro Collector Spotter

When moving the Agilent 1200 Series micro collector/spotter inside the laboratory, no special precautions are needed. However, if the micro collector/spotter needs to be shipped to another location via carrier, ensure:

- ✓ The transport assembly is in the park position. Use the ChemStation or the Control Module for this command.

To move the arm to the park position:

- 1 Switch to the **Diagnosis** view of the ChemStation and select **Fraction Collector > Maintenance Positions.** from the **Maintenance** menu
 - 2 In the upcoming dialog box click **Park Arm.**
- ✓ The vial tray and the sample transport mechanism is secured with the transport protection foam.



2 Configuration and Operation of the Micro Collector/Spotter

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Introduction

The Agilent 1200 Series micro collector/spotter G1364D is an instrument for micro scale fraction collection or MALDI Spotting with the Agilent 1200 Series capillary or nanoflow LC systems. With these systems the collection of very low volume fractions requires

- low delay volumes.
- high position accuracy of the capillary tip.
- liquid contact control for reproducible collection of small fractions without carryover.

The principles of operation for the Agilent 1200 Series micro collector/spotter will be illustrated in this chapter along with detailed descriptions on how to set up the method and change the configuration. This will enable the user to run the instrument with optimal performance.

The Agilent 1200 Series micro collector/spotter designed for low flow rates from below 100 $\mu\text{l}/\text{min}$ down to 100nl/min. In order to keep dispersion a single low volume PEEK coated fused silica capillary is used to direct the flow to the vessel or MALDI target. A permanent waste position is built in to allow automated and continuous collection of the eluent.

The proper alignment of the capillary tip is crucial for precise collection of the fractions, especially if 384 conical-well plates or MALDI targets are used. A sensor pad for automated control of the capillary tip position was therefore integrated into the micro collector/spotter. A diagram of the fraction collector, which indicates the most important components, is shown in [Figure 1](#) on page 35

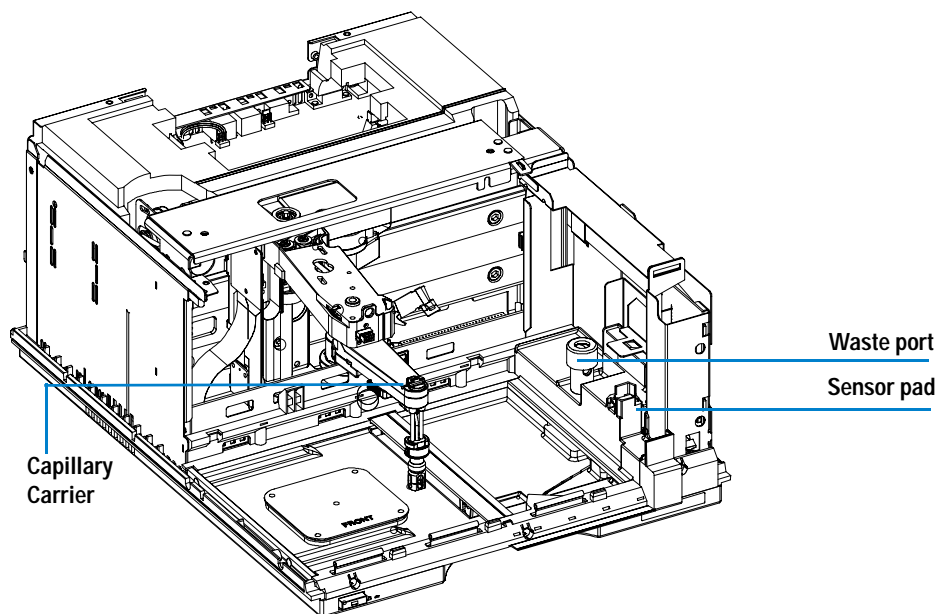


Figure 1 The Agilent 1200 Series Micro Collector/Spotter

In order to ensure reproducible fraction collection and MALDI spotting without cross contamination and an accurate control of the collected volumes the *liquid contact control* mode as illustrated in [Figure 2](#) on page 36 has been developed. During the entire collection process the instrument ensures that the droplet at the capillary tip is in constant contact with the surface of the well, the MALDI plate or the liquid being collected. At the start of fraction collection the droplet is delivered at the bottom of the well or the MALDI plate. While the well is filled the capillary tip moves continuously upwards, keeping in constant contact with the liquid surface. At the end of fraction collection, the capillary tip moves sharply upwards in order to cut the contact between the liquid and the capillary. During the collection the flow rate and the geometry of the well are taken into account to calculate the current fill level.

2 Configuration and Operation of the Micro Collector/Spotter

Introduction

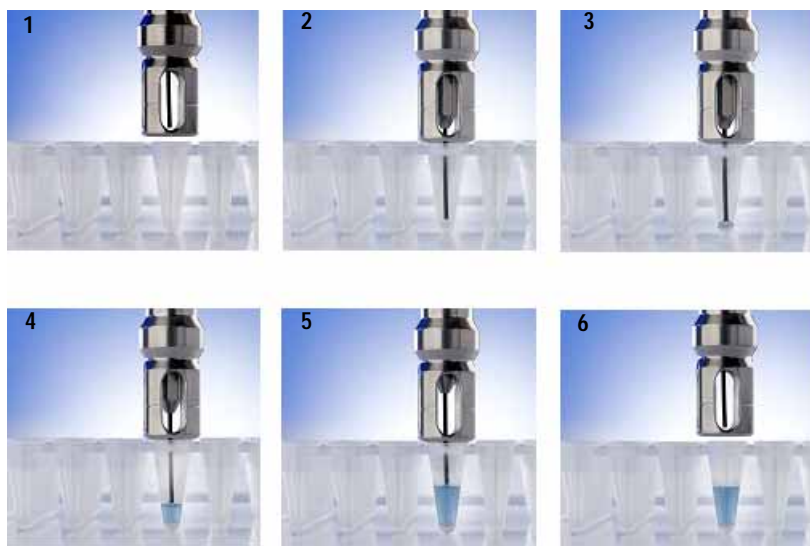


Figure 2 Liquid Contact Control

Initialization and Reset

After power-on the instrument delays the initialization for approximately two minutes to vent the fraction compartment. Then the door lock is activated and the micro collector/spotter remains in the *not ready* condition (*yellow*) waiting for the manual reset.

Before you reset the micro collector/spotter the door has to be closed. To reset the instrument click on the micro collector/spotter icon in the graphical user interface of the ChemStation and start the **Reset**. If the main pump is switched ON, a dialog is displayed, which reminds the user to switch OFF the pump. Then the reset procedure is started without the risk of contaminating well plates or fractions.

During the initialization process the vertical and the horizontal position of the capillary tip is determined. The micro collector/spotter performs the capillary check by pressing the capillary tip onto the sensor pad. After this capillary check a message is displayed, if any further capillary alignment is required. For more details about capillary alignment refer to [“Choosing and Installing the Micro Collector/Spotter Capillary”](#) on page 38.

Switching the micro collector/spotter lamp

The micro collector/spotter includes a lamp to illuminate the fraction compartment. The lamp is specifically required for the position accuracy calibration as described on [page 100](#) and [page 102](#).

The operation of the lamp requires firmware revision A.05.09 or higher and ChemStation revision A.10.02 or higher. To activate the lamp switch open the command line in the ChemStation and type:

microafclampon to switch the lamp ON
microafclamppoff to switch the lamp OFF

Configuration of the Micro Collector/Spotter

Choosing and Installing the Micro Collector/Spotter Capillary

The micro collector/spotter is delivered with three capillaries each having a different inner diameter: 25, 50 and 100 μm . The 50 μm capillary is installed in the micro collector/spotter, whereas the 25 and the 100 μm capillaries are in the accessory kit (see “[Micro Collector/Spotter Accessory Kit](#)” on page 137).

To optimize the micro collector/spotter for different flow ranges the correct capillary has to be selected and installed.

For micro fraction collection we recommend:

- flow rates below 4 $\mu\text{L}/\text{min.}$: the yellow 25 μm i.d. capillary G1364-87304 (delay volume app. 0.25 μL)
- flow rates between 4 and 30 $\mu\text{L}/\text{min.}$: the green 50 μm i.d. capillary G1364-87305 (delay volume app. 1 μL)
- flow rates between 30 and 100 $\mu\text{L}/\text{min.}$: the black 100 μm i.d. capillary G1364-87306 (delay volume app. 4 μL)

For MALDI spotting we recommend:

- for overall flow rates (LC flow + matrix flow) < 8 $\mu\text{L}/\text{min.}$: the green 50 μm i.d. capillary G1364-87305 (delay volume app. 1 μL)
- for overall flow rates (LC flow + matrix flow) > 8 $\mu\text{L}/\text{min.}$: the black 100 μm ID capillary G1364-87306 (delay volume app. 4 μL)

The exchange procedure is described in “[Replacing Micro Collector/Spotter Capillary Assembly](#)” on page 117. An Exchange of the fraction collector capillary might also be necessary, if the capillary is bent or blocked by particles.

Adjusting the peakwidth of your detector

When using capillaries in these flow ranges for peak-based fraction collection a **Peakwidth** of 0.1 min. (**Responsetime** < 2s) or less should be selected to ensure a fast signal processing. Otherwise the compound might already be flushed through the micro collector/spotter, when the peak is detected. If a smaller peakwidth is set the flow rate ranges can be increased.

Capillary Alignment for micro fraction collection

During the initialization process the vertical and the horizontal position of the capillary tip is determined. The micro collector/spotter facilitates the capillary check by pressing the capillary onto the sensor pad. Then the relative capillary length as well as the degree to which the needle is bent is determined. 384-well plates require a higher position accuracy than 96-well plates. Consequently a slightly bent capillary might be tolerated for 96-well plates, whereas 384-well plates require the installation of a new capillary or a position accuracy calibration as described in [“Position Accuracy Calibration for 384-Well Plates”](#) on page 100. After the capillary check procedure a message is displayed, if any further capillary alignment is required.

CAUTION

If solvents with high concentrations of matrix, buffer or salt are used, the capillary has to be flushed thoroughly with salt free water after analysis. Such a procedure prevents the capillary and the waste port from clogging.

The capillary length has to be adjusted between the following limits (see [Figure 3](#)):

- For 384-conical-well plates (PCR plates) where high position accuracy is required the capillary tip should be approximately set to the groove at the center of the well-plate adapter.
- When 96-well plates or deep 384-well plates are used the capillary tip must be set to the lower edge of the well-plate adapter opening in order to reach the bottom of the wells.

2 Configuration and Operation of the Micro Collector/Spotter

Configuration of the Micro Collector/Spotter

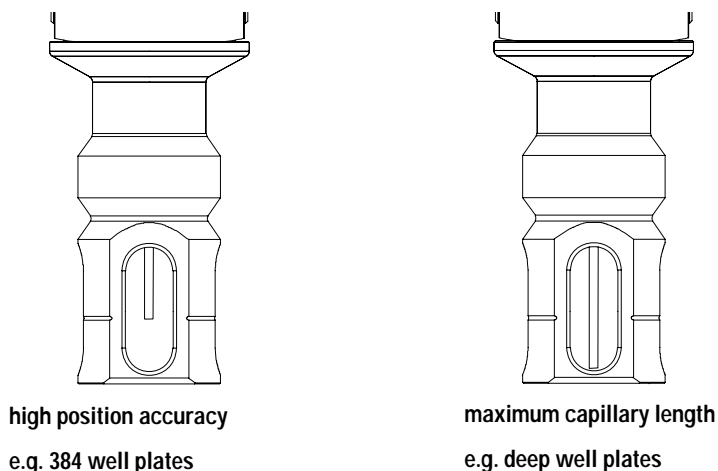


Figure 3 Positions of the capillary tip

Capillary Alignment for MALDI spotting

If the micro collector/spotter is used for MALDI spotting, the well-plate adapter has to be replaced with the spotting adapter (G1364-83205), which is supplied in the accessory kit (see [Table 19](#) on page 137).

For MALDI Spotting applications we recommend using the MALDI spotting tip. This tip prevents that the droplet from flowing upwards on the outer capillary wall.

- 1 Remove all trays and move the fraction collector arm to the **Change Parts** position (see [“Maintenance Functions”](#) on page 93).
- 2 Turn the well-plate adapter 90 degrees and remove it from the capillary carrier arm.
- 3 Slide the spotting adapter onto the capillary carrier arm and turn it 90 degrees to fixate it. The capillary length should be aligned as displayed in [Figure 4](#).
- 4 Attach the MALDI spotting tip to the capillary. The MALDI spotting tip should be aligned to from a flat surface with the capillary tip. For correct alignment, leave the spotting tip in the plastic strip and push the spotting tip onto the capillary.

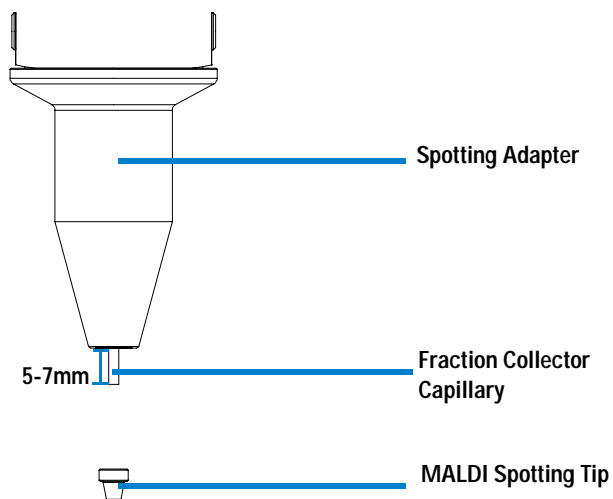


Figure 4 Capillary Alignment for MALDI Spotting

Configuration of the ChemStation

To setup or change the configuration parameters of your fraction collector select **More Fraction Collector > Configuration** from the Instrument menu or right-click on the fraction collector icon in the graphical user interface. In the Fraction Collector Configuration dialog box (Figure 5) the configuration of the Trays, the Fraction Delay Volumes, the Collection Order, the Collection Order, the Needle Movement and the well-plate/MALDI target type can be specified.

Fraction Collector Configuration : Instrument 1

Trays
Tray A: 4 WellPlates
Tube Volume [ml]: 0.00
Tube Height [mm]: 0

Fraction Delay Volumes

Detector	Volume[ml]
----------	------------

Collection Order

☒ Flow by row
☐ Column by column
☐ Shortest path by column
☐ Shortest path by row

Collection Mode

☒ Discrete fractions
☐ Continuous flow

Needle Movement

☐ into location Depth: 5.0 mm
☐ above location Distance: 2.0 mm
☒ contact control Distance: 0.2 mm

Well-Plates

Plate Type: 96EppendorfC ☐ Treat as 4x96

Installed Plates

<input checked="" type="checkbox"/> Plate 1 (front left)	<input checked="" type="checkbox"/> Plate 3 (front right)
<input checked="" type="checkbox"/> Plate 2 (back left)	<input checked="" type="checkbox"/> Plate 4 (back right)

Reserved Locations

Row(s):
Column(s):
Single Location(s):

OK Cancel Help More >>

Figure 5 Micro Collector/Spotter Configuration dialog

Trays

In the online ChemStation the configuration of the Trays is recognized automatically. In the offline ChemStation the tray configuration can be chosen in a dropdown menu.

Fraction Delay Volumes

The fraction delay volumes specify the volume between the detector cell and fraction collector capillary tip. This volume is required to calculate the time delay between the detection of the peak in the detector and the start of the collection in the fraction collector. For the micro collector/spotter G1364D this volume depends on the installed capillaries. The fraction delay volume can be calculated by:

$$\text{fraction delay volume} = \text{volume (detector outlet capillary)} + \text{volume (fraction collector inlet capillary)}$$

The volumes of the most common Agilent 1200 Series detector outlet capillaries and the fraction collector inlet capillaries are summarized in [Table 1](#) on page 43 and [Table 2](#) on page 44. To identify the capillary check the label for the part number, internal diameter and capillary length. For time-based operation the detector cell can be bypassed to minimize the delay volume.

Table 1 Detector outlet capillaries

description	part number	length/mm	ID/ μm	Volume/nl
DAD/MWD outlet capillary for 500 nl flow cell (pre mounted)	G1315-87338	120	100	945
DAD/MWD outlet capillary for 500 nl flow cell (alternative)	G1315-87328	120	50	235
DAD/MWD outlet capillary for 500 nl flow cell (pre mounted, old flow cell kit)	G1315-68708	700	75	3100
DAD/MWD outlet capillary for 80nl flow cell (pre mounted)	G1315-87328	120	50	235
DAD/MWD outlet capillary for 80nl flow cell (alternative)	G1315-87318	600	25	300

2 Configuration and Operation of the Micro Collector/Spotter

Configuration of the Micro Collector/Spotter

Table 2 micro collector/spotter inlet capillaries

description	part number	length/mm	ID/ μ m	Volume/nl
micro collector/spotter capillary 25 μ m ID (alternative)	G1364-87304	500	25	250
micro collector/spotter capillary 50 μ m ID (pre mounted)	G1364-87305	500	50	1000
micro collector/spotter capillary 100 μ m ID (alternative)	G1364-87306	500	100	4000

NOTE

Due to the tolerance for the capillary ID, small adjustments in delay volume might be necessary in order to optimize the performance of your system.

Collection Order

The **Collection Order** describes capillary movement during fraction collection. Four different strategies are possible: row-by-row and column-by-column either in one direction or in two directions.

Collection Mode

Selection of the **Collection Mode** is not available for the micro collector/spotter and therefore greyed out in this configuration. The micro collector/spotter always operates in the **continuous flow** mode.

Needle Movement

Into location In the into location mode the capillary tip moves into the well to the specified **Depth** (in mm).

Above location In the above location mode the capillary tip stays at the specified **Distance** (in mm) above the well during fraction collection.

Contact Control In this mode the capillary tip moves down until it reaches the specified distance between capillary tip and well bottom. This ensures that the forming droplet is in contact with the well bottom. During the continuing filling process the capillary tip moves upwards while staying in contact with the surface of the collected liquid.

For low flow rates ($< 1 \mu\text{L}/\text{min}$) a small **Distance** ($< 0.2 \text{ mm}$) should be chosen. This value should be increased to $0.3\text{--}0.7 \text{ mm}$ in particular for MALDI spotting at higher flow rates.

NOTE

The capillary movement in the Contact Control mode depends on the flow rate that is delivered by the Agilent 1200 Series pump. If your LC system contains more than one pump, the flow rate of the first pump in your system determines the needle movement. In this case verify in your system configuration (**Instrument > Configure 1200 access**) that this pump is the first one in the **Configured Modules** list. In addition this pump should also be selected as main pump (**Instrument > Change main pump**).

Well Plates

In the **Well Plates** section the type of well plates or MALDI targets used in a well plate tray can be configured. The well plate type used can be chosen from the Plate Type dropdown menu. More detailed information about all preconfigured well plates and MALDI targets can be found in the Instrument menu. A list of recommended Agilent well plates can be found in [Table 12](#) on page 130 and a list of supported MALDI targets is available in [Table 16](#) on page 133. To configure other well plates or MALDI targets choose **Configure Well Plates** from the **Instrument** menu.

NOTE

Note the orientation of the well plate or the MALDI plate. The starting position A1 is always at the back left corner of the plate.

Only one type of well plate can be used on the well plate tray.

The checkbox **Treat as 4 x 96** is only available for 384 well plates and allows splitting the 384 well plate virtually into 4 separate 96-well plates. This only changes the collection order, but the numbering of the wells remains as indicated on the plate. The filling order of each quarter is as specified in the Collection Order section. When the 384 well plate is split into four equal quarters the order of the four plates is the same as displayed in the Installed Plates section. Each tray position that contains a well plate has to be checked.

2 Configuration and Operation of the Micro Collector/Spotter

Configuration of the Micro Collector/Spotter

In the **Reserved Locations**, you have the possibility to specify locations that will not be used for Fraction Collection (see [Table 3](#)).

Table 3 Syntax for the definition of Reserved Locations

Location	Syntax	Description
Rows	A	Row A can't be used
	A,B	Rows A and B can't be used
	A-D	A, B, C and D can't be used
	A-D,F	Rows A, B, C, D and F can't be used
Columns	1	Column 1 can't be used
	1,2	Columns 1, and 2 can't be used
	1-4	Columns 1, 2, 3 and 4 can't be used
	1-4,12	Columns 1, 2, 3, 4 and 12 can't be used
Single locations	G12,H12	Locations G12 and H12 can't be used

Setting up a Micro Collector/Spotter Method

To setup the method parameters of the micro collector/spotter select **Setup Fraction Collector** from the Instrument menu or right-click on the fraction collector icon in the graphical user interface. This will open the Setup Fraction Collector dialog box as displayed in [Figure 6](#). In the Setup Fraction Collector dialog box general method settings are specified.

Set up Fraction Collector : Instrument 1

Fraction Trigger Mode
☐ Off ☒ Use Timetable ☐ Peak-based
 Max. Peak Duration: 1.0 min

Peak Detectors

Detector	Working Mode	Up Slope [Unit/s]	Down Slope [Unit/s]	Threshold [Unit]	Upper Threshold [Unit]	Unit
1 DAD1	Off	5.00	5.00	5.000	3000.000	mAU
2 UIB	Off	5.00	5.00	5.000	3000.000	mV

Fraction is collected when a peak is detected by:
☒ all selected peak detectors ☐ at least one selected peak detector
☐ Use MSD for Mass-based Fraction Collection

Timetable

	Time	Trigger Mode	Max. Peak Duration [min]	# of Fractions	Timeslices [min]	Contact Adj.
1	0.00	Off				1.0
2	2.50	Time-based		10		1.0
3	10.00	Off				

Buttons: Insert, Append, Cut, Copy, Paste

Time
 Stop time: as Pump no Limit min
 Post time: Off min

Auxiliary
☒ Recovery on the track
 Contact Control Adjustment: 1.0

Auxiliary
 Max. fill volume per location as configured: pl
☐ Store temperature signal
☐ Store delay sensor signal
☐ Store UIB signal

Buttons: OK, Cancel, Help

Figure 6 Set up Fraction Collector dialog box

Fraction Trigger Mode

The Fraction Trigger Mode can either be **Use Timetable** or **Peak-based** can be selected. Furthermore for the Peak-based mode a time for **Maximum Peak Duration** can be specified. When the Peak-based mode is chosen, it overrules all

settings in the Timetable. If the LC system is used for analytical chromatography without fraction collection or MALDI spotting, the micro collector/spotter can be switched **Off**.

CAUTION

The spot size for MALDI spotting is specified by the flow rate and the collection time. If peak-based trigger mode is selected for MALDI spotting, the collection time and therefore the spot size is flexible. In this case we recommend specifying a maximum fill volume per location as described in [“Auxiliary”](#) on page 50.

Peak Detectors

The **Peak Detectors** section in this screen comprises a list of all Peak Detectors connected to the system. As working modes for each peak detector **Threshold only**, **Threshold/Slope** and **Slope only** are possible. In the Threshold only mode the settings for Up Slope, Down Slope and Upper Threshold in the subsequent columns are ignored. Fraction collection is triggered whenever the detector signal exceeds the specified threshold value. When the signal drops below the threshold value fraction collection is stopped. In the Slope only mode fraction collection is triggered on the slopes of the detector signals. Adequate values for Up Slope and Down Slope can be specified in the corresponding fields. In the Threshold/Slope mode fraction collection is triggered on the corresponding values for threshold and slope. For more detailed information concerning working with threshold and slopes please refer to Agilent’s Application Note “Sophisticated peak-based fraction collection – working with up and down slope” publication number 5988-7895EN.

The option to specify an **Upper Threshold** becomes important, if the absorbance exceeds the linear range of the UV-Vis detector. At high absorbance values the light intensity on the detector is extremely low and consequently detector noise will superimpose the detector signal. In this case the detector noise might lead to wrong trigger commands for the micro collector/spotter. As soon as the detector signal exceeds the **Upper Threshold**, settings for slopes will be ignored until the signal drops again below the Upper Threshold.

When using more than one peak detector fraction collection can be triggered either when **all selected peak detectors** detect a peak or when **at least one selected peak detector** detects a peak based on the settings in the detector table.

The **Timetable** can be used to program changes in the Fraction Trigger Mode during the analysis by entering a time in the Time field and appropriate values in the fields of the timetable. The trigger modes can be specified as Off,

Peak-based and Time-based. Time ranges can be defined for the different trigger modes In the column headed **Time**. If the Off mode is selected, no fractions are collected.

Whenever the Peak-based mode is specified fractions will be collected based on the peak detection parameters given in the Peak Detector table. Additionally a Maximum Peak Duration in minutes has to be specified. This parameter can be used to stop the fraction collection in cases where the baseline drifts, and the signal does not drop below the specified threshold value. The limits are from 0.1 min. to 9999.0 min. This parameter is mandatory if you use Peak Controlled fraction collection, but is disabled for Time Based fraction collection.

Timetable

When the **Time-based** mode is chosen two different options are available:

- A specific number of fractions or time slices can be specified. The **# of Fractions** refer to the number of fractions that are collected in the corresponding time frame in the table. This parameter is applicable only for time-based fraction collection. **The maximum spotting rate is 20 spots/min.**
- Alternatively time slices can be set-up that define the collection time for each fraction. This parameter is only available in the time-based fraction collection mode. Based on the maximum spotting rate of 20 spots per minute **the minimum time slice for one spot is 3s.**
- **Contact Adj** is a parameter, which is only required for MALDI Spotting. For details please read "[Contact Control Adjustment](#)" on page 50.

To edit the Timetable, the functions **Insert**, **Append**, **Cut**, **Copy** and **Paste** are available.

To access the additional sections in the Setup Fraction Collector dialog box click the **More** button.

Recovery

If the **Recovery on the Track** mode is checked, the solvent that is delivered between the fractions/spots is collected on the same plate for recovery. The capillary carrier will move to the next well following collection order, e.g. row-by-row.

In the Data Analysis view as well as in the Report a fraction list indicates the wells that contain fractions and the wells that contain recovery. With this information the user is able to recover compounds that weren't collected.

Contact Control Adjustment

This parameter is only required for MALDI Spotting. If you are adding your matrix online as described in “[Online Matrix Delivery](#)” on page 59, the flow rate at the capillary tip of the micro collector/spotter is higher than the flow rate that is measured at the flow sensor of your Agilent 1200 Series capillary or nanoflow pump. Consequently the needle movement in the **Contact Control** mode has to be adjusted. The factor can be calculated by

$$\text{Contact Control Adjustment} = (\text{pump flow} + \text{matrix flow}) / \text{pump flow}$$

Some examples are presented in the [Table 4](#) below.

Table 4 Examples for Contact Control Adjustment

Pump Flow/ nl	Matrix Flow/ nl	Contact Control Adjustment
500	0	1.0 (default)
500	500	2.0
1000	500	1.5
500	1000	3.0

The Contact Control adjustment can also be used to optimize the MALDI spotting process and if the flow is splitted before the fraction collector.

Auxiliary

In the Auxiliary section the **Maximum fill volume** per location can be specified. If **as configured** is selected, the pre-configured volume (see **Instrument > Pre-configured Wellplate Types**) is used. This ensures that the location (well, vial or tube) cannot be overfilled during fraction collection. This volume can be further reduced by defining a customized volume.

By default all MALDI plates are configured with an infinite fill volume. The **Maximum fill volume** can be used to determine a maximum spot size, if the peak-based trigger mode is used.

Additional check boxes in this section provide the opportunity to **Store** the **temperature signal** and the **UIB signal**.

Fraction Preview

To determine the appropriate fraction collection parameters the Agilent ChemStation provides a valuable tool that becomes accessible by selecting the button labelled Fraction Preview Tool (Figure 7) in the Peak Detectors section.

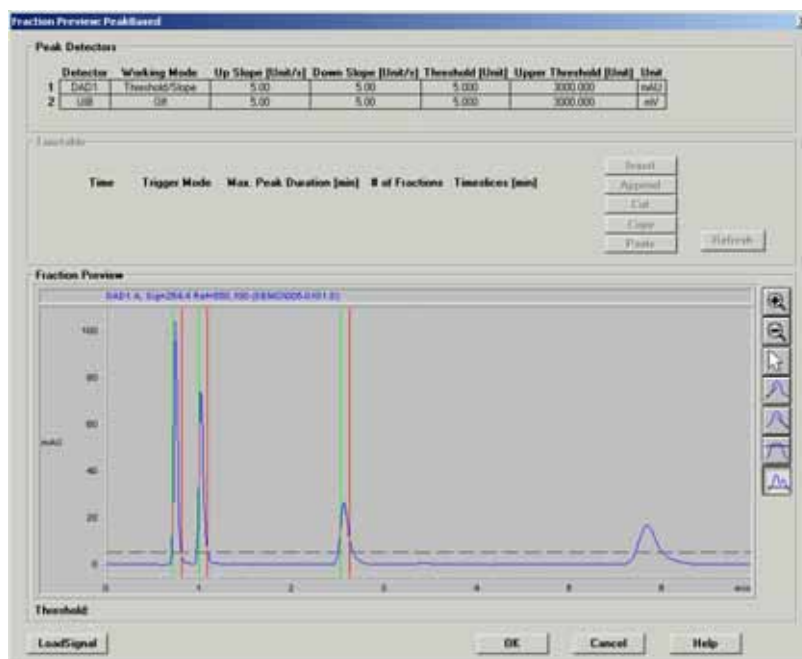


Figure 7 Fraction Preview dialog box

The Fraction Preview screen allows testing the fraction collection parameters against a sample chromatogram. It can also be used to optimize the fraction collection parameters interactively. With the help of this tool, values for up and down slope as well as for upper and lower threshold can easily be graphically specified. A chromatogram, e.g. a pilot run, can be loaded by pushing the **Load Signal** button. Parameters can now be changed either

2 Configuration and Operation of the Micro Collector/Spotter

Setting up a Micro Collector/Spotter Method

manually in the Detector Table and Timetable or graphically in the Fraction Preview screen. By pushing the desired buttons on the right hand side of the Fraction Preview screen the chromatogram can be zoomed, the values for up and down slope can be specified and the upper and lower threshold level can be set-up. The graphically specified values are automatically transferred to the Peak Detector Table.

Starting your run with fraction collection or MALDI spotting

Resetting the current fill levels

To protect the collected fractions/spots for contaminations and to avoid overfilling of single positions the Agilent 1200 Series micro collector/spotter stores current fill levels on the module and blocks used positions. If a tray with new well plate or MALDI target is inserted in the micro collector/spotter, all fill levels can be reset by removing and re-installing the 4-well-plate tray (not just the well plate or MALDI plate carrier). After re-installation of the tray a pop-up window as displayed in [Figure 8](#) will be displayed. Click on **Yes** to reset all fill levels from the previous run.



Figure 8 Reset fill volumes

CAUTION

The window ([Figure 8](#)) to reset current fill levels is not displayed, if the well plate or the MALDI target is removed without removing the complete tray. Consequently contaminated fractions, unwanted overfill events or split fractions might be observed during the following LC run.

NOTE

The run time for a system with a fraction collector must be extended by the delay time, to ensure the complete collection of all compounds.

The run time is calculated as:

$$\text{run time} = \text{end of last peak} + \text{delay time} \\ (\text{with delay time} = \text{delay volume} / \text{flow rate})$$

This calculation is only required, if the flow rate or the delay volume (capillary) is changed. For further info read "[Fraction Delay Volumes](#)" on page 43.

Assignment of Start Location

The start location for fraction collection or MALDI spotting can either be assigned in the **Sample Info** (Figure 9) dialog box in the RunControl menu, in the **Sequence Parameters** (Figure 10) dialog box or in the **Sequence Table** in the Sequence menu. All fraction start locations are entered in the format

micro collector/spotter # - plate number - row - column, e.g. 1-P1-A-01

with

micro collector/spotter # = 1,2; plate number = P1-P4; row is A to H and column = 1-12 for 96-well plates or the Agilent AP-MALDI plate (for different well plates or MALDI plates refer to the description from the plate manufacturer).

NOTE

Note the orientation of the MALDI plate. The starting position A1 is always in the back left corner of the plate.

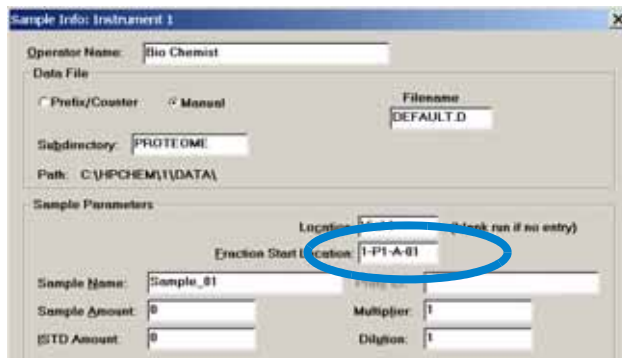


Figure 9 Sample Info dialog box

All settings made in the Sequence Table overrule the settings chosen in the Sequence Parameters screen. In addition to the exact position for fraction collection start in the Sequence Table it is also possible to specify Next Plate and Next Location. Then the fraction collection is started at the next free plate or at the next free location.

If no fraction start location has been specified, the fraction collection starts at first empty location on the well plate or MALDI target.

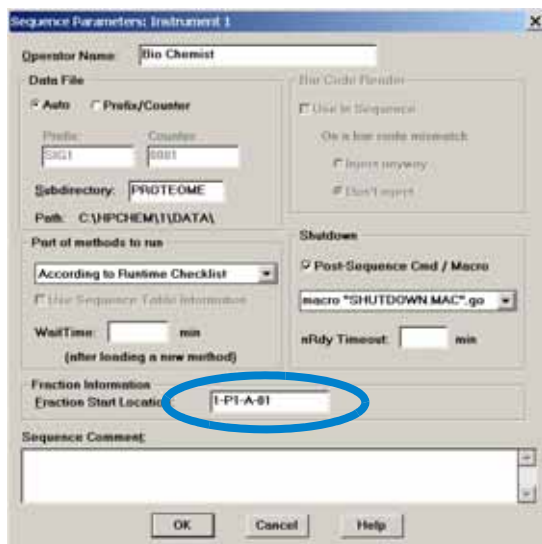


Figure 10 Start Location in Sequence Parameters dialog box

To start a single run

- click on the Start button in the graphical user interface of the ChemStation.
- select **Run Method** from the **RunControl** menu.
- press F5

To start the sequence

- click on the Start button in the graphical user interface of the ChemStation.
- select **Run Sequence** from the **RunControl** menu.
- press F6

Online tick marks

To display tick marks for fraction start and stop events in you Online Plot click on the **Change** button in your **Online Plot** window. Then check **Show fraction collection ticks** in the **Edit Signal Plot** window (Figure 11).

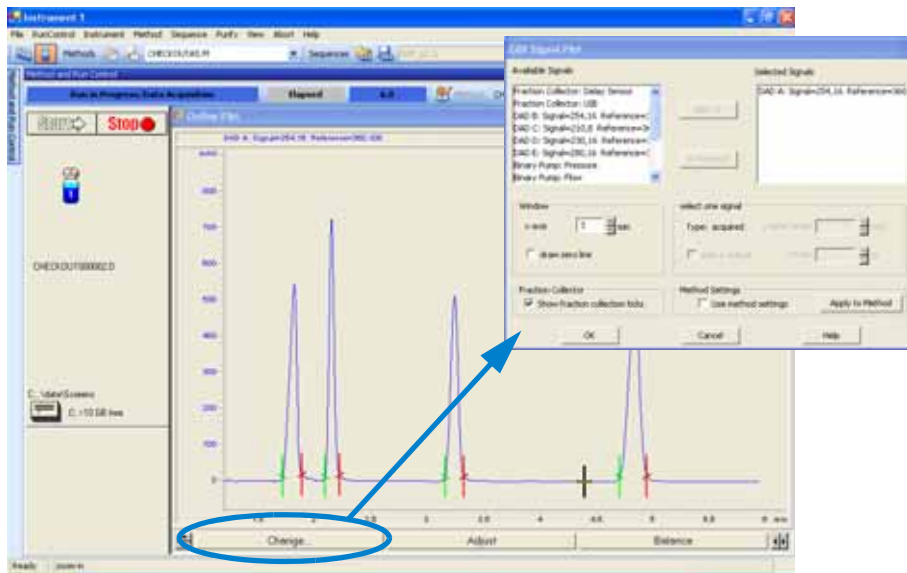


Figure 11 Online Tick Marks

Viewing your Results

Data Analysis

In order to display the tick marks for the collected fractions or spots on the screen, click on **Signal options** from the **Graphics** menu. Then choose **Separated** in the Layout drop-down menu.

To review your chromatograms, file information and a fraction list, select the Data Analysis view from the drop-down menu and press the Fraction Task button as displayed in [Figure 12](#).

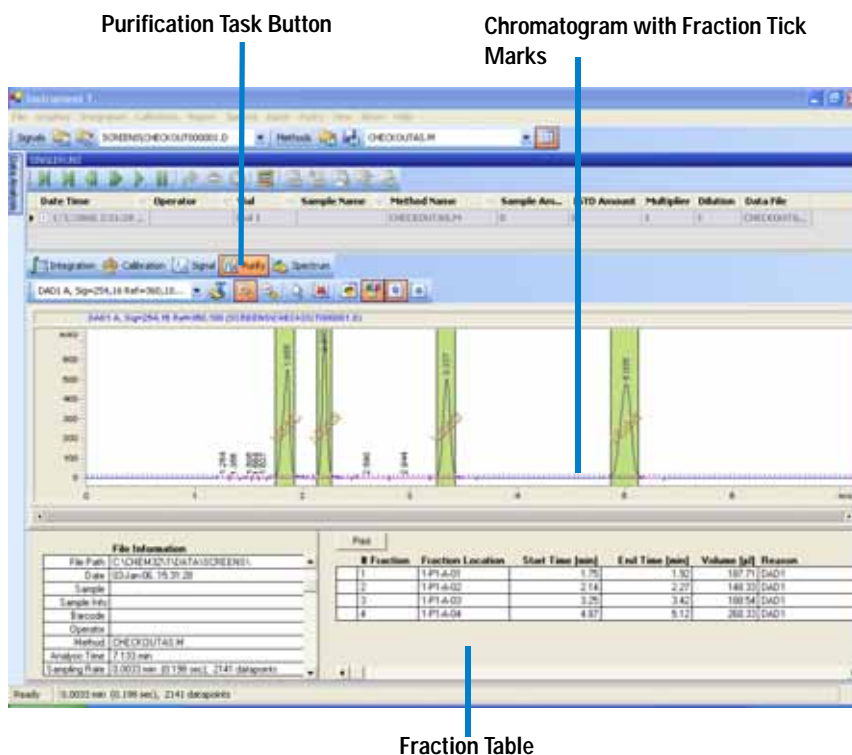


Figure 12 Data Analysis View

Report

In order to create reports with a fraction table and tick marks the **Specify Report** dialog box the item **Add Fraction Table and Tick Marks** has to be checked.

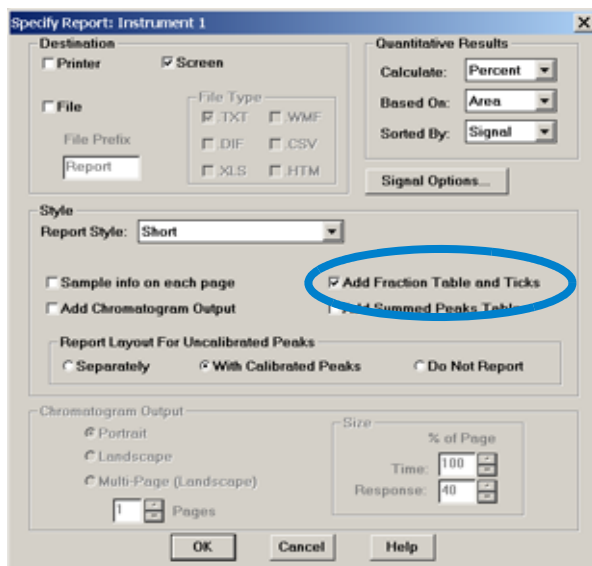


Figure 13 Fraction collection report setup

Online Matrix Delivery

The Agilent 1200 Series micro collector/spotter can be configured for online matrix delivery. For this application the Online Matrix Kit (G1364-68706) is required (see Table 15 on page 131). This kit contains all accessories required to control a syringe pump and to connect the syringe to the LC flow path. An overview is displayed in Figure 14.

NOTE

For low flow rates and for MALDI spotting with online matrix delivery, we recommend using the plug to close the waste container instead of the waste tubing. The plug (G1364-26105) is included in the micro collector/spotter accessory kit.

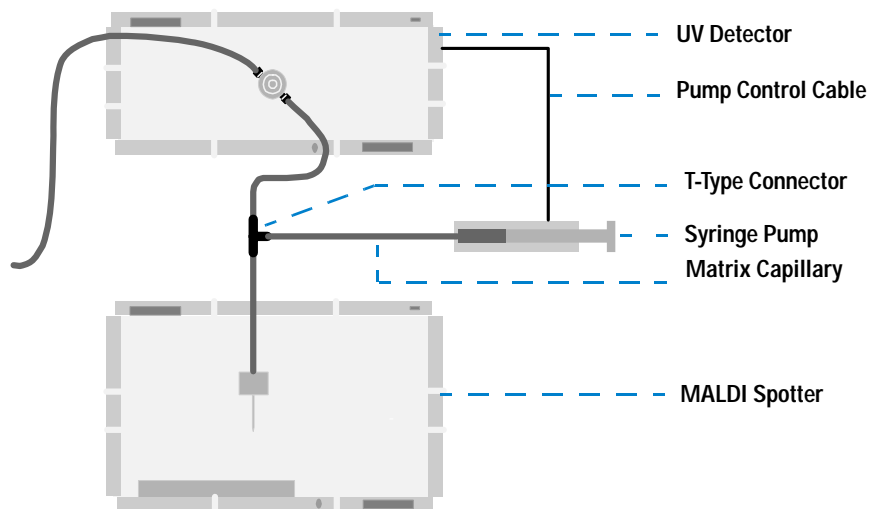


Figure 14 Online Matrix Delivery

Syringe Pump Control

The Online Matrix Kit contains an External Contact Board BCD (G1351-68701) to control the syringe pump. In addition a cable to connect the KDS 200 syringe pump is provided (5181-1536). To connect other syringe pumps a

general purpose control cable (18594-60520) can be ordered and configured. The BCD board can be installed in any Agilent 1200 Series LC module. For installation details refer to the corresponding service manual.

The pump control can easily be setup in the ChemStation as displayed in [Figure 15](#). To open the dialog box select the module where the external contact board is installed and click on the icon in the graphical user interface. Then select **Contacts**.

The default setting should be set as **open**, i.e. the syringe pump is switched off. In the example that is shown in [Figure 15](#) the syringe pump is switched off by default. Using the Timetable the matrix flow can be switched as required during the run. We recommend switching the syringe pump on at least 10 minutes before the start of the LC run in order to prime the matrix capillary and ensure that the matrix is added to the LC flow immediately after the run is started.

Contact 1 is **open**: syringe pump is switched off

Contact 1 is **closed**: syringe pump is switched on

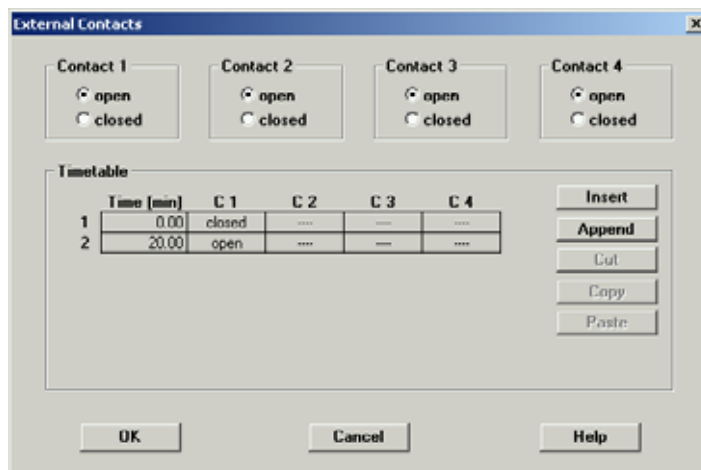


Figure 15 Syringe Pump Control through External Contacts

Capillaries and Fittings

The Online Matrix Kit contains:

- peek coated fused silica capillary 125 μ m ID, 550mm length (G1375-87318)

- T-type connector (5042-8519)
- Connector syringe to capillary (5042-8517)

To setup the flow connections the outlet capillary from the UV detector, the fraction collector capillary and the capillary from the matrix pump have to be connected to the T-type Connector as displayed in [Figure 14](#) on page 59.

We recommend using

- the 50 µm ID fraction collector capillary (G1364-87305) for overall flow rates (LC flow and matrix flow) < 8 µl/min.
- the 100 µm ID fraction collector capillary (G1364-87306) for overall flow rates (LC flow and matrix flow) > 8 µl/min.

The peek coated fused silica capillary (125 µm ID, 550mm length) is used to deliver the matrix solution from the syringe to the T-type connector, where it is merged with the flow from the Agilent 1200 Series LC system.

The maximum spot size strongly depends on the type of the MALDI plate and can range between 0.5 and 5 µl. The flow rate has to be adjusted to optimize the performance with spotting rate of 20 spots/min:

$$\text{maximum flow rate} = \text{maximum spot size} \times 20 \text{ spots/min}$$

Table 5 Examples for maximum flow rates

maximum spot size	maximum spotting rate	maximum flow rate (LC flow + matrix flow)
0.5 µl	20 spots/min	10 µl/min
1 µl	20 spots/min	20 µl/min
2 µl	20 spots/min	40 µl/min
5 µl	20 spots/min	100 µl/min

Contact Control Adjustment

If the matrix is added online, the flow that is delivered by the Agilent 1200 Series capillary or nanoflow pump is different from the flow at the capillary tip in the MALDI spotter. To account for this additional flow the Contact Control Adjustment has to be specified as described on [page 50](#).

2 Configuration and Operation of the Micro Collector/Spotter

Online Matrix Delivery

CAUTION

If solvents with high concentrations of matrix, buffer or salt are used, the capillary has to be flushed thoroughly with salt free water after analysis. Such a procedure prevents the capillary and the waste port from clogging.

Check-out procedures

The purpose of the check-out injection is the verification of the correct installation and configuration of the Agilent 1200 Series micro collection/spotting system. In addition the checkout procedure can be used for troubleshooting, if the Agilent 1200 Series micro collector/spotter doesn't operate as expected.

The checkout procedure depends on the application and is different for micro fraction collection into well plates and for MALDI spotting.

Check-out procedure for micro fraction collection

The check-out procedure is based on the procedure for the Agilent 1200 Series capillary LC system. After the check-out sample (01080-68704) has been injected, four peaks have to be detected and four fractions collected.

Prerequisites

- ✓ The Agilent 1200 Series capillary LC system has been installed and prepared as described in the Agilent 1200 Series capillary LC system manual G1376-90011.
- ✓ The Agilent 1200 Series micro collector/spotter has been installed and configured. The delay volume has been specified.
- ✓ A well plate is installed as plate 1 in the 4 well plate tray. If 384 well plates are used, the [“Arm Length Calibration”](#) on page 97 and the [“Position Accuracy Calibration for 384-Well Plates”](#) on page 100 have to be executed before running the check-out.
- ✓ All wells are empty and the fill levels, which are stored on the module, have been reset. To verify this remove and insert the 4 well plate tray. Then click on **Yes** in displayed dialog box.

Table 6 Method parameters for micro fraction collection check-out

Agilent 1200 Series modules	<ul style="list-style-type: none"> • micro degasser • capillary pump (20µl flow sensor) • micro autosampler/ micro well plate sampler • column compartment (optional) • diode array detector/ multiple wavelength detector with 500nl or 80 nl flow cell • micro collector spotter
Column	ZORBAX SB C18, 5 µm, 150 x 0.5mm (5064-8256)
Sample	Check-out sample (01080-68704); diluted 1:10 in Acetonitrile
Flow	15 µl/min
Stoptime	10 minutes
Solvent A	30% (HPLC grade water)
Solvent B	70% (HPLC grade acetonitrile)
Wavelength DAD/MWD	Signal A: 254/4nm, Reference: 360/80nm
Peakwidth (Responsetime)	> 0.1min (2s)
Injection volume	200 nl
Column temperature	25.0 °C or ambient
Fraction trigger mode	Peak-based
Max. peak duration	1 min
DAD/MWD working mode	Threshold/slope
Upslope	5 mAU/s
Downslope	5 mAU/s
Threshold/Upper threshold	5 mAU/ 3000 mAU

Expected result

Run a single injection under the conditions as described in [Table 6](#). You should observe a chromatogram with four peaks and four fractions should have been collected in the well plate.

Check-out procedure for MALDI spotting

To check-out the MALDI spotting functionality a run with blank injection with time-based spotting is performed to verify that Agilent 1200 Series micro collector spotter consistently collects onto the MALDI target.

Prerequisites

- ✓ The Agilent 1200 Series capillary LC or nanoflow system has been installed and prepared as described in the Agilent 1200 Series capillary LC system manual G1376-90011 or the Agilent 1200 Series nano LC system quick start guide G2226-90011.
- ✓ The Agilent 1200 Series micro collector/spotter has been installed and configured for MALDI spotting. The delay volume has been specified. The MALDI adapter and the MALDI spotting tip have been installed.
- ✓ The “Position Accuracy Calibration for MALDI Targets” on page 102 has been executed before running the checkout.
- ✓ A MALDI plate carrier with supported MALDI target is installed as plate 1 in the 4 well plate tray.

Table 7 Method parameters for MALDI spotting check-out

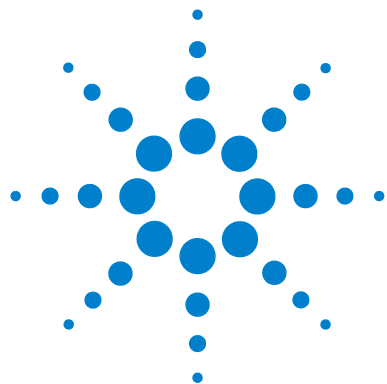
Agilent 1200 Series modules	<ul style="list-style-type: none"> • micro degasser • capillary pump (20µl flow sensor) or nanoflow pump • micro autosampler/ micro well plate sampler • column compartment (optional) • diode array detector/ multiple wavelength detector with 500nl or 80 nl flow cell (optional) • micro collector/spotter (configured for MALDI Spotting)
Column	ZORBAX SB C18, 5 µm, 150 x 0.5mm (5064-8256) for the capillary pump ZORBAX SB C18, 3.5 µm, 150 x 100µm (5065-9910) or similar column for the nanoflow pump
Sample	blank injection (no sample required)
Flow	5 µl/min (capillary pump) or 1 µl/min (nanoflow pump)
Stoptime	3 minutes (capillary pump); 6 minutes (nanoflow pump)
Solvent A	30% (HPLC grade water)

Table 7 Method parameters for MALDI spotting check-out (continued)

Solvent B	70% (HPLC grade acetonitrile)
Injection volume	blank injection
Column temperature	25.0 °C or ambient
Fraction trigger mode	Use Timetable
1 With the capillary pump	
Time	0.5 minutes
Trigger mode	Time-based
# of fractions	20
Time	2.5 minutes
Trigger mode	Off
2 With the nanoflow pump	
Time	0.5 minutes
Trigger mode	Time-based
# of fractions	10
Time	5.5 minutes
Trigger mode	Off

Expected result

Run a single injection under the conditions as described in [Table 7](#). You should observe 20 spots with the capillary LC system or 10 spots with a nanoflow LC system. The spots should be consistently aligned on the MALDI target. Due to evaporation the spots collected earlier during the run will be smaller than the later spots.



3

Control Module Screens of the Fraction Collector

Major keys on the Agilent 1200 Series Control Module 68

Screens available from the Analysis view 69

Screens available from the System view 80

This chapter is intended to introduce the user to the functionality for operation of the Agilent 1200 Series micro collector/spotters G1364D with the Agilent 1200 Series control module.

This chapter will provide an overview of the control module features. Please use the User Manual of the Agilent 1200 Series control module or the **i**-key (online help) for further details.

To use the full functionality of the Agilent 1200 Series micro collector/spotter the Agilent ChemStation is required. With the Agilent 1200 Series control module:

- The alignment tools for arm length calibration and position accuracy alignment are not available.
- Only one fraction start location per sequence can be specified
- No tick marks are displayed in the online plot.
- The flexibility of managing data and reporting is very limited.

Nevertheless the Agilent 1200 Series control module can be helpful during the operation of the micro collector/spotter. For example the user will be able to trigger the fraction collector manually or specify the flow rate of a non-Agilent 1200 Series pump.



Major keys on the Agilent 1200 Series Control Module

ESC	Return to previous screen, abort any change of parameters and toggle between the last two top layer views
m	Open context sensitive menus
i	Information/Help
Enter	Store changed parameters or execute a choice
Done	(If available) Activate settings of current screen
On/Off	Switch on individual Instrument(s) or complete System
Start	Start a location range or sequence
Plot	View online signals
Views	Change between analysis - (samples)- status - system views

NOTE

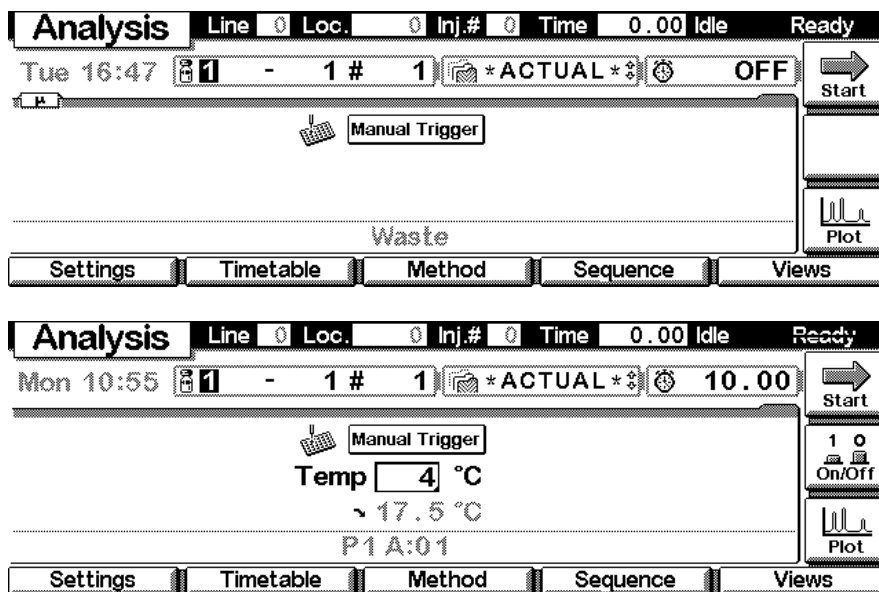
The screens shown on the next pages are based on:
Control Module firmware revision B.04.0x (G1323B)
HPLC Module firmware revision A.05.06 and higher

NOTE

In case the control module's display seems to be frozen (hang-up due to a communication problem on the CAN bus), unplug the control module from the HPLC module and reconnect.

Screens available from the Analysis view

Analysis view This is the start-up screen, if the Agilent 1200 Series micro collector/spotter or thermostatted micro collector/spotter is the only configured Agilent 1200 Series LC module.

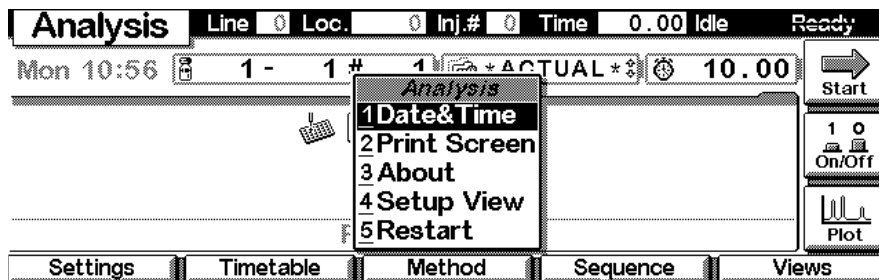


On/Off (F7) allows you to turn on the Thermostat Module.

The **m**-key allows access to the context sensitive menus. **Date&Time** allows you to change time settings. **Print Screen** sends the screen to a configured printer location. **About** tells you the current firmware revision and the serial# of your control module. **Setup view** leads you to analysis view configuration for additional Agilent 1200 Series LC modules. **Restart** re-boots the control module. If multiple instruments are configured in the Analysis screen the context menu contents will vary, depending on the cursor position, when the m-key is pressed (depending on highlighted module).

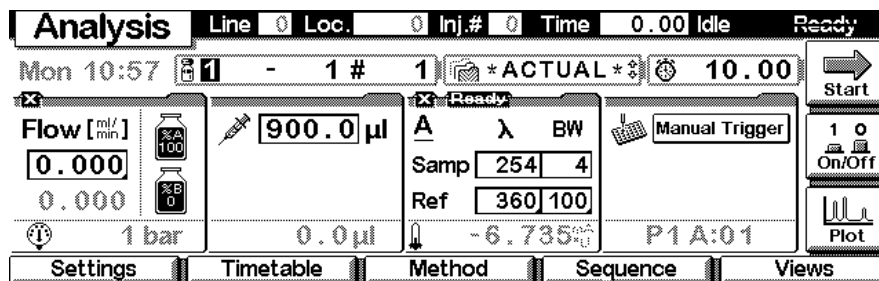
3 Control Module Screens of the Fraction Collector

Screens available from the Analysis view



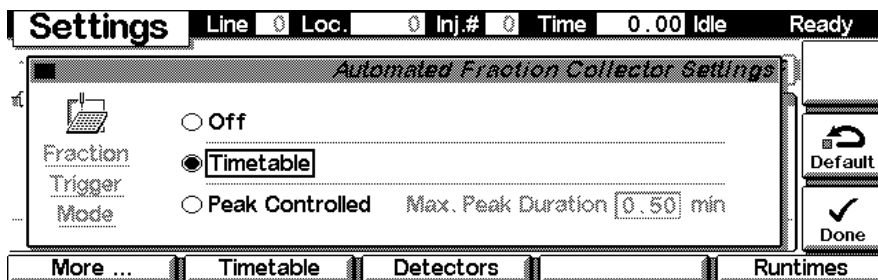
In the Setup view, modules can be added or removed to the analysis view. Select a module, you want to move, with the help of the **selection**-keys. Use function keys **F7/F8 (Remove/Add)** to move the highlighted module. Changes must be activated with **Done (F6)**.

Here, e.g. the Agilent 1200 Series pump, the injector and the detector parameters are shown on the display as well. The number of parameters for each module is reduced as additional modules are added to the display. A maximum of 4 modules can be shown simultaneously. If more modules are connected to the system, you have to choose 4 of them in **Setup view**.

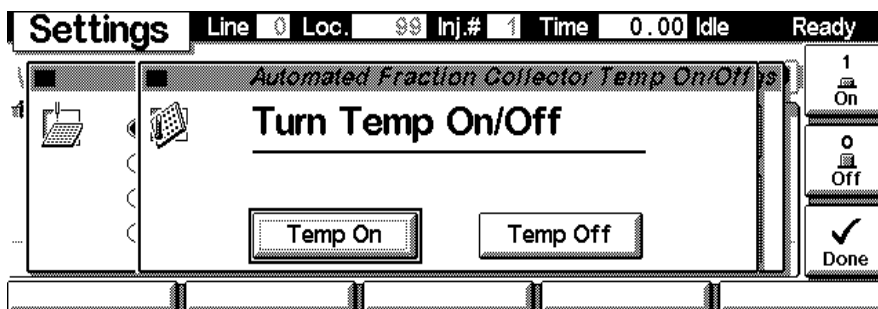


Settings With the **F1**-key (**Settings**) you open a pull-down menu where you can select the micro collector/spotter module. Within **Settings** you can change the type of fraction collection (**Off**, **Timetable**, or **Peak Controlled**). There are different sets of parameters available through the **F1-3** and **F5** navigation-keys for setting path operation of the micro collector/spotter. **F7 (Default)** resets the micro collector/spotter to default values. Changes must be acknowledged with **F6 (Done)**.

Settings



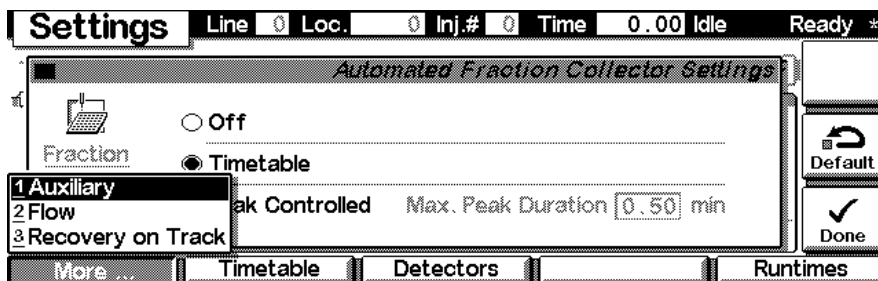
Settings screen with thermostat connected. **On/Off (F8)** allows you to switch the thermostat on or off.



Use the **m**-key for the context sensitive menu. The **Status** command pulls up a module specific screen displaying details of configured trays. **Reset** will initialize the micro collector/spotter.

Settings More...

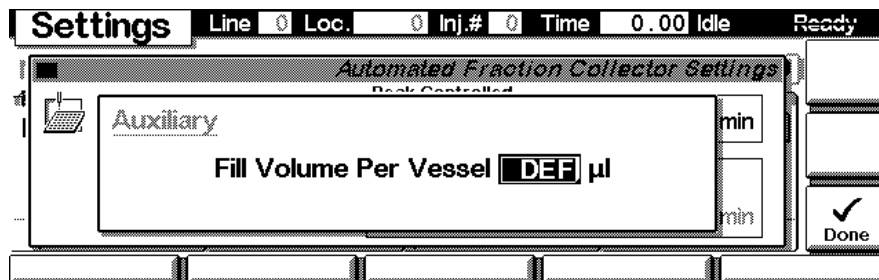
The **F1**-key (**More**) opens a pull-down menu.



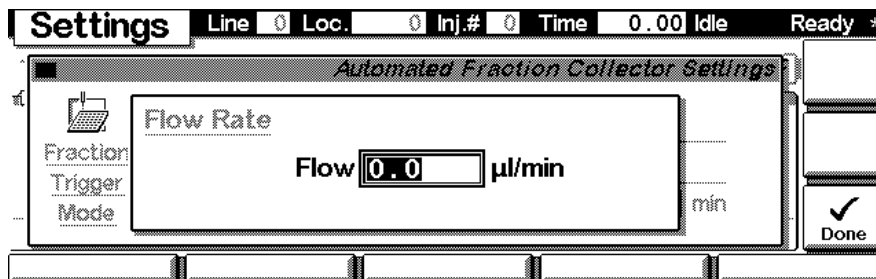
3 Control Module Screens of the Fraction Collector

Screens available from the Analysis view

By selecting **More...Auxiliary** on the pull-down menu you can enter the micro collector/spotter **Fill Volume Per Vessel** for the currently installed well plates or MALDI targets. Leaving this value at the “DEF” value will completely fill the vessels according to their specified size. Changes must be acknowledged with **F6 (Done)**.

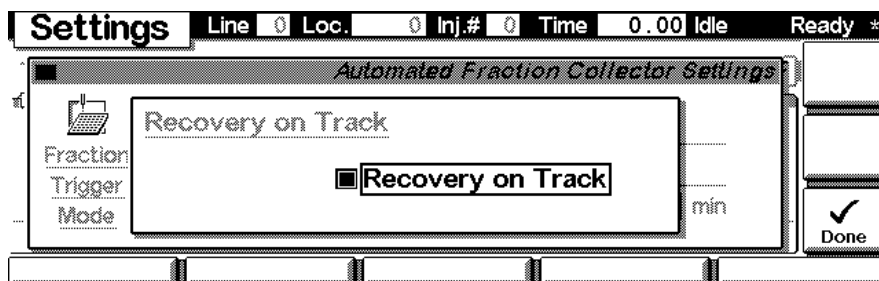


By selecting **More...Flow** you can specify the flow rate if no Agilent 1200 Series pump is configured in LC system. The flow rate is required to calculate the needle movement in the liquid contact control mode and to calculate delay time between peak detection and collection of the fraction. With an Agilent 1200 Series pump the flow rate will be automatically transferred to the micro collector/spotter. Changes must be acknowledged with **F6 (Done)**.



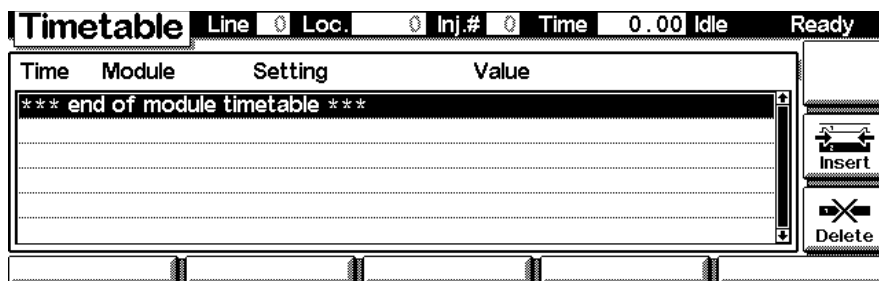
By selecting **More...Recovery on Track** on the pull-down menu you can activate the recovery mode of the micro fraction collector/spotter. With **Recovery on Track** selected the module will collect recovery fractions into the well plate, if no peak is detected. As soon as the next fraction is triggered the

fraction collector will move to the next position. With this functionality no eluent is flushed to waste and the sample can be recovered from the well plate, if it hasn't been correctly collected. **Recovery on Track** is recommended for valuable samples and if the chromatographic behavior of the sample is unknown, e.g. during method development. Changes must be acknowledged with **F6 (Done)**.



Settings -
Timetable

With the **F2-key (Timetable)** in the **Settings** screen you can list the timetable for the micro collector/spotter. Press **F7-key (Insert)** to edit the selected entry or **F6-key (Delete)** to remove the selected/highlighted entry.



Edit each entry line according to your needs, by entering a time for a Setting and by selecting a Setting with the help of the pull-down menu. Use **F7 (Enter)** to insert the current settings. Changes must be acknowledged with **F6 (Done)**.

3 Control Module Screens of the Fraction Collector

Screens available from the Analysis view

Timetable Line 0 Loc. 0 Inj.# 0 Time 0.00 Idle Ready *

Time [min] Module Setting

2.00 AF Collector Trigger Mode

Mode Time 0.50 min

fields change with setting selected

Enter

Done

Timetable Line 0 Loc. 0 Inj.# 0 Time 0.00 Idle Ready *

Time	Module	Setting	Value
2.00	AF Collector	Trigger Mode	0.50 min each fraction
10.00	AF Collector	Trigger Mode	Off
*** end of module timetable ***			

Insert

Delete

Use the **m**-key for the context sensitive menu. It gives you additional tools for the timetable.

Timetable Line 0 Loc. 0 Inj.# 0 Time 0.00 Idle Ready *

Time	Module	Setting	Value
*** end of module timetable ***			

Timetable

- 1 Copy Line
- 2 Paste Line
- 3 Print Timetable

Insert

Delete

Settings - Detectors With the **F3-key (Detectors)** in the **Settings** screen you can configure and **Enable (F8)** or **Disable (F7)** any detector that is part of a system for peak detection. Changes must be acknowledged with **F6 (Done)**.

Detector	Use	Upslope	Downslope	Threshold
DA Detector	✓	OFF	OFF	5.000

☐ Use MSD for Mass-based Fraction Collection

Fraction is collected, when

☐ all Peak Detectors detect a peak

☒ at least one Peak Detector detects a peak

Enable
Disable
Done

Settings - Thermostat With the **F4-key (Thermostat)** in the **Settings** screen you can set the micro collector/spotter thermostat temperature. Changes must be acknowledged with **F6 (Done)**.

Automated Fraction Collector Settings
Peak Controlled

Temperature °C 26.8 °C min

☐ at Power-On turn temperature on min

On/Off
Done

3 Control Module Screens of the Fraction Collector

Screens available from the Analysis view

Settings - Run times

With the **F5**-key (**Runtimes**) in the **Settings** screen you can change the stop time and the post-run time for the selected module individually. Changes must be acknowledged with **F6** (**Done**).

The screenshot shows the 'Settings' screen with a title bar containing 'Line 0 Loc. 0 Inj.# 0 Time 0.00 Idle Ready'. The main area is titled 'Automated Fraction Collector Settings' and 'Peak Controlled'. It displays 'Stoptime' as 'OFF' with '0.00 min' and 'Posttime' as 'OFF' with '0.00 min'. Below these is a note: 'overrides HPLC system runtime settings'. On the right side, there are three buttons: a top button, a middle button, and a bottom button labeled 'Done' with a checkmark icon.

Method screens

Use **Esc** to return to the **Analysis** screen. Use the **F3**-key (Method) to view the parameters in a method. Use the **F8**-key (**Save As**) to save the method in the module(s). The PC-Card-key is only active when a PCMCIA card is inserted in the control module. **Module** (**F1**) and **Method** (**F2**) give access to methods stored on the Agilent 1200 Series LC module or the PC-Card. **Timetable** (**F3**) gives access to the timetable of a method (if specified).

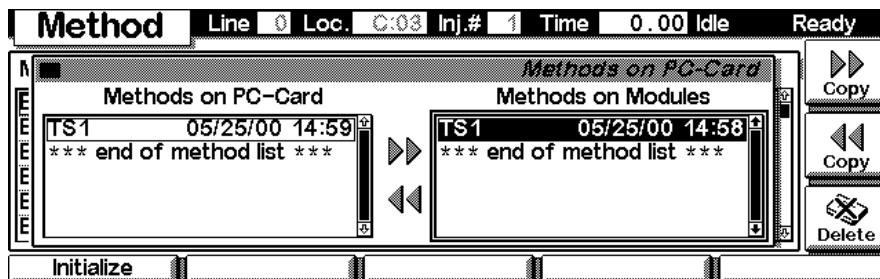
NOTE

The PCMCIA card must have been inserted into the control module prior to the last start-up in order to be recognized.

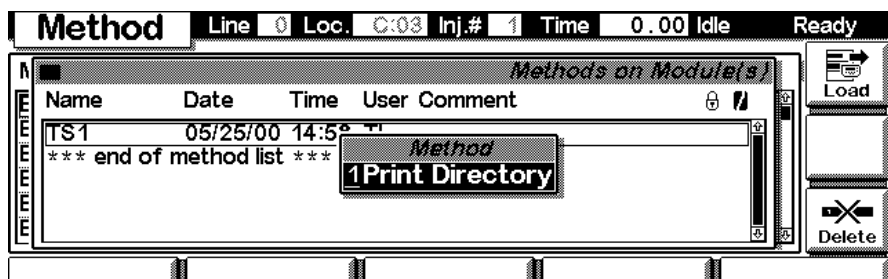
The screenshot shows the 'Method' screen with a title bar containing 'Line 0 Loc. 0 Inj.# 0 Time 0.00 Idle Ready *'. The main area is a table with 'Setting' and 'Value' columns. The settings are: 'Stoptime' (OFF), 'Posttime' (OFF), 'Trigger Mode' (Timetable), 'Flow' (10.0 ml/min), and 'Recover on track' (On). The table ends with '*** end of method ***'. On the right side, there are three buttons: 'Save As' (with a floppy disk icon), 'Default' (with a circular arrow icon), and 'Print' (with a printer icon). At the bottom, there are four buttons: 'Module', 'PC-Card', 'Timetable', and 'Edit'.

Setting	Value
Stoptime	OFF
Posttime	OFF
Trigger Mode	Timetable
Flow	10.0 ml/min
Recover on track	On
*** end of method ***	

After pressing **F1** or **F2** use the **Right/Left** selection keys to switch between PC-Card and Instrument window. Use the **Up/Down** selection keys to select the method. Use **F7/F8 (Copy)** to copy a selected method from one window to the other one. Use **F6 (Delete)** to delete a selected method.



Press the **m**-key to open the context sensitive menu for printing the method directory.



3 Control Module Screens of the Fraction Collector

Screens available from the Analysis view

sequence In the sequence view (F4 from the **Analysis Screen**) you can specify the sequence's **End Actions** (F1), load sequences from **PC-Card** (F2) (if present) or proceed to a **method** table (F3). F6 (**Delete**) and F7 (**Insert**) allow you to edit lines in the sequence table.

Sequence Line 0 Loc. 99 Inj.# 1 Time 0.00 Idle Ready

Line	From	To	#	Inj. Vol	Method	Wait	Calibration
***	end of sequence ***						

Start
Insert
Delete

End Actions PC-Card Method Logbook

Sequence Line 0 Loc. 99 Inj.# 1 Time 0.00 Idle Ready

End Actions

On end of ...
■ Sequence

Timeout OFF min

☐ Load Method * ACTUAL *
☒ Turn off Nothing
☐ Turn off complete HPLC system

Done

Parameters that can be entered into a line in the sequence table.

Sequence Line 0 Loc. 99 Inj.# 1 Time 0.00 Idle Ready

Sequence Insert Line 1

Vial 1 to 1 #Inj. 1 Vol DEF µl

Method * ACTUAL * Wait after loading 0.00 min

☒ use Calibration to apply calibration settings

Enter
Done

Calibration

Fractions From the Analysis View press the **F5**-key (**Views**) and select **Fractions**.

The screenshot shows the 'Analysis' screen with the following details:

- Header:** Line 0, Loc. 99, Inj.# 1, Time 0.00, Idle, Ready.
- Time/Date:** Wed 09:10.
- Sample Info:** 99 - 100 # 1, *ACTUAL*, 1.00.
- Flow:** 0.000 ml/min.
- %B:** 0.0.
- Inj Vol:** 0.0 µl.
- Pressure:** 0 bar.
- Menu:**
 - 1 Analysis
 - 2 Samples
 - 3 **Fractions** (highlighted)
 - 4 Status
 - 5 System
- Buttons:** Settings, Timetable, Method, Sequence, Views.

This is an example if an Agilent 1200 Series micro collector/spotter is configured stand-alone. Here you can define a **Start** position for a fraction, select a collection **Scheme**, choose a **Trigger Mode** etc.

The screenshot shows the 'Fraction' screen with the following details:

- Header:** Fraction Loc. Waste, Time 0.00, Idle, Ready.
- Start at:** P1 A:01.
- Scheme:** Selection of collection schemes using icons.
- Trigger Mode:** Timetable.
- Plot Area:** Four sub-plots showing data trends.
- Buttons:** Manual Trigger, Timetable, Method, Views.

3 Control Module Screens of the Fraction Collector

Screens available from the System view

Screens available from the System view

System view Use the **Esc**-key until you receive **Views** on the **F5**-key. Choose **System** from the pull-down menu. This screen shows the last activities in the system.

The screenshot shows the 'Analysis' screen with the following details:

- Line: 0, Loc.: 99, Inj.#: 1, Time: 0.00, Idle, Ready
- Wed 09:10, 99 - 100 # 1, *ACTUAL*%, 1.00
- Flow: 0.000 ml/min, Inj Vol: 0.0 µl
- %A: 100, %B: 0
- Pressure: 0 bar, 99%, 0.0 µl
- Wavelength: ~69.496
- Buttons: Settings, Timetable, Method, Sequence, Views
- Right side menu: 1 Analysis, 2 Samples, 3 Fractions, 4 Status, 5 System (selected), 6 Off

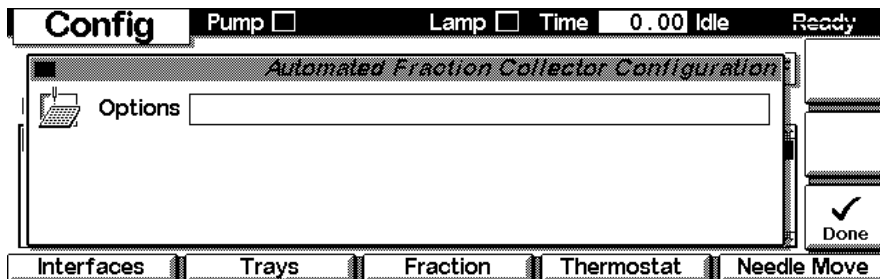
System - Configure Use the **Esc**-key to receive **Views** on the **F5**-key in the Analysis Screen. Choose **System** from the pull-down menu. Use the **F2**-key (**Configure**) to select the micro collector/spotter. Here you define further special set points for the micro collector/spotter operation. Changes must be acknowledged with **F6** (**Done**).

The screenshot shows the 'System' screen with the following details:

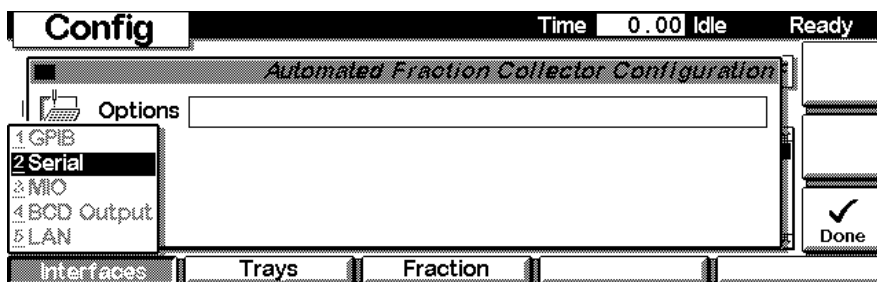
- Time: 0.00, Idle, Ready
- Tue 16:48, EMF
- Buttons: Start, Plot
- Table of messages:

Module	Message	Id	Date	Time
AF Collector	Front door closed	STATE	10/25	16:47:16
AF Collector	Front door open	STATE	10/25	16:47:00
AF Collector	Needle is nearly centeric	STATE	10/25	16:46:51
AF Collector	1 HPLC System 4 Plates	INFO	10/25	16:46:06
AF Collector	2 AF Collector reset	INFO	10/25	16:45:10

- Buttons: Control, Configure (selected), Tests, Records, Views

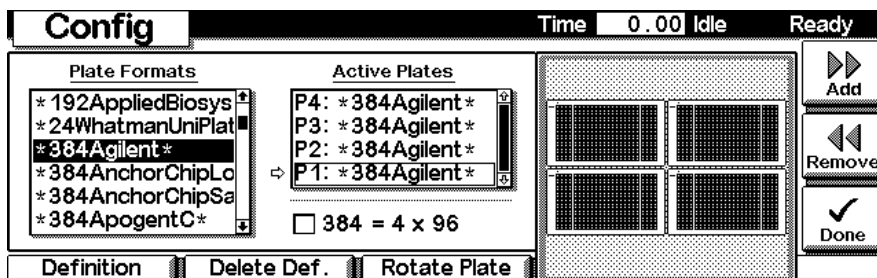


Use the **F1**-key (**Interfaces**) to access the interface settings (if required). Changes must be acknowledged with **F6** (**Done**).



Configure - trays

Use the **F2**-key (**Trays**) to view and configure the (automatically) detected sample trays. Different views will come up depending on the installed trays. The standard tray for the micro collector spotter is the 4 well plate tray



Select/highlight a tray configuration with the help of the selection keys. **F7/F8** (**Add/Remove**) allows you to move configurations from available Plate

3 Control Module Screens of the Fraction Collector

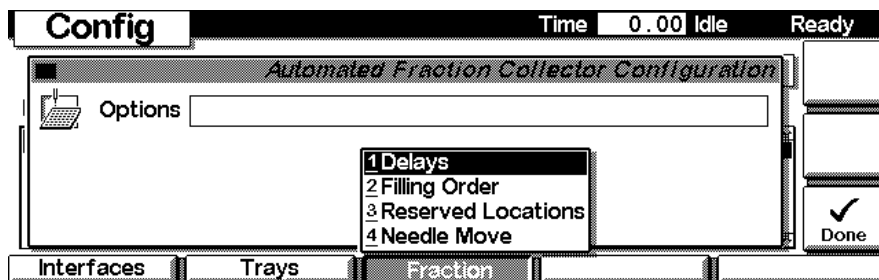
Screens available from the System view

Formats to the Active Plates and vice versa. Press **F2 (Delete Def.)** to delete a user defined definition. Press **F3 (Rotate Plate)** to rotate the selected plate. Use **F1 (Definition)** to edit the user defined definition of a well-plate. Changes must be acknowledged with **F6 (Done)**.

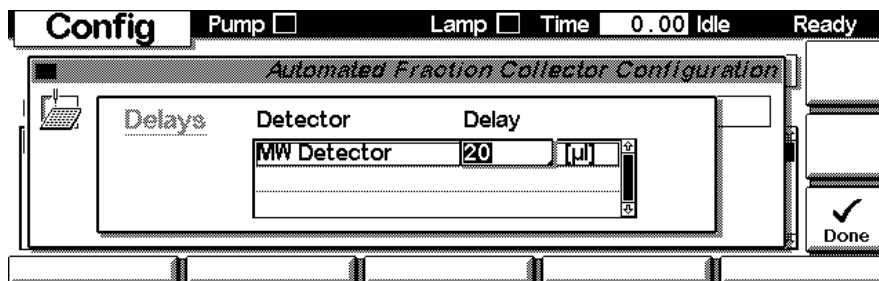
NOTE

Only one type of well plate or MALDI target can be selected at the same time.

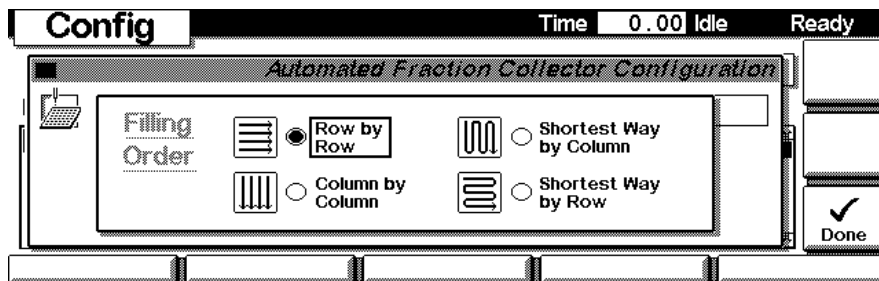
Configure - fraction **F3 (Fraction)** in the main Configuration screen allows you to access the **Delay** volume definition screen, to specify **Filling Order** and **Reserved** or **Needle Movement**.



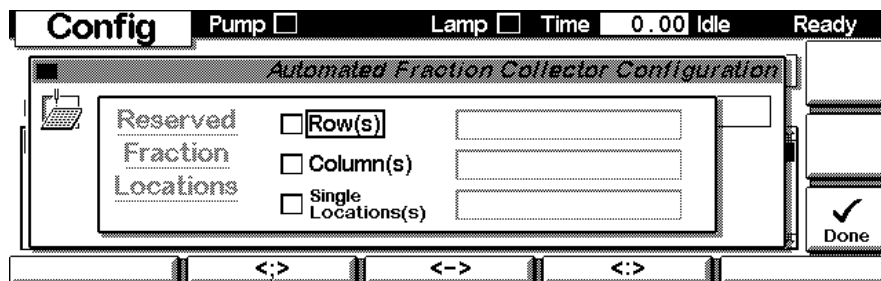
Configure - fraction - delays **Delays** in the Fraction Configuration screen allows you to configure the delay volume between each connected detector and the micro collector/spotter. **Edit (F8)** allows to enter new values for the selected detector. Changes must be acknowledged with **F6 (Done)**.



Configure - fraction - filling order **Filling Order** in the Fraction Configuration screen allows you to configure the filling order of the trays in the micro collector/spotter. **Edit (F8)** allows to enter new values for the selected detector. Changes must be acknowledged with **F6 (Done)**



Configure - reserved locations **Reserved Locations** allows you to specify reserved locations by **Rows**, **Columns** or **Single Locations**, that are not used (filled) during the collection. Changes must be acknowledged with **Done (F6)**.

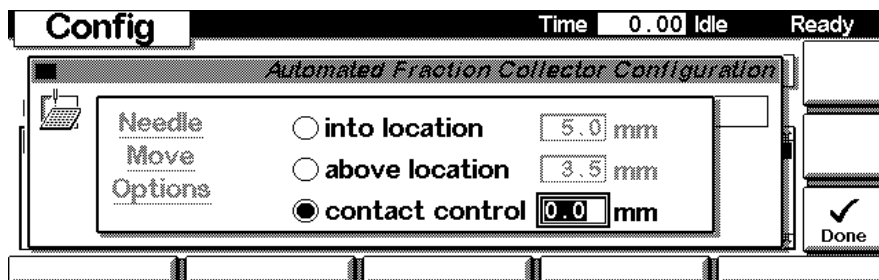


Configure - Needle Movement The **Needle Movement** allows you to specify the position of the needle (capillary) tip during fraction collection. With **into location** the capillary tip moves to the specified depth in the well. With **above location** the capillary tip stays at the specified distance above the well. For micro fraction collection or MALDI spotting we recommend to use the **contact control** mode. In this mode the capillary tip moves to the specified distance above the bottom of the well or the MALDI target. When the fraction is collected the needle will move upwards while staying in contact with the liquid surface of the collected fraction. This movement types enables the

3 Control Module Screens of the Fraction Collector

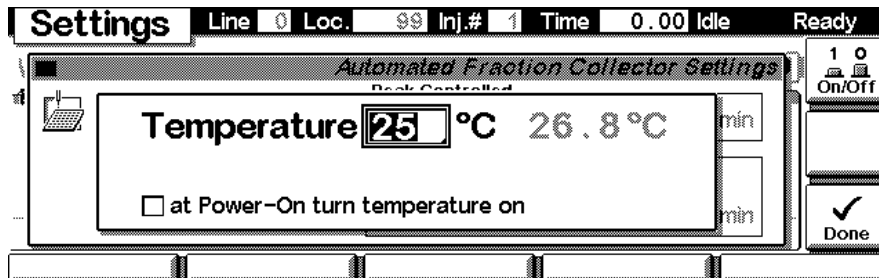
Screens available from the System view

Agilent 1200 Series micro collector/spotter to collect low volumes without carry-over or air bubbles at the bottom of the well. Changes must be acknowledged with **Done (F6)**.



Configure -
Thermostat

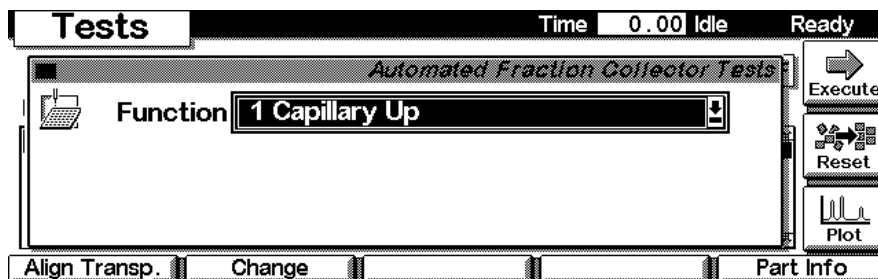
F4 (Therm) in the main Configuration screen allows you to configure the temperature and turn on conditions of the thermostatted micro collector/spotter. Changes must be acknowledged with **F6 (Done)**.



System - tests

Press **F3** from the **System** View and choose the micro collector/spotter to access the micro collector/spotter **Tests** screen.

The Tests screen gives you access to multiple tests and features such as the automatic transport unit **Alignment (F1)** tool, maintenance procedures to **Change (F2)** parts on the micro collector/spotter. **Parts Info (F5)** provides information about the type and revision of the installed transport unit assembly. The pull-down menu in the center of the screen gives you access to various step commands.



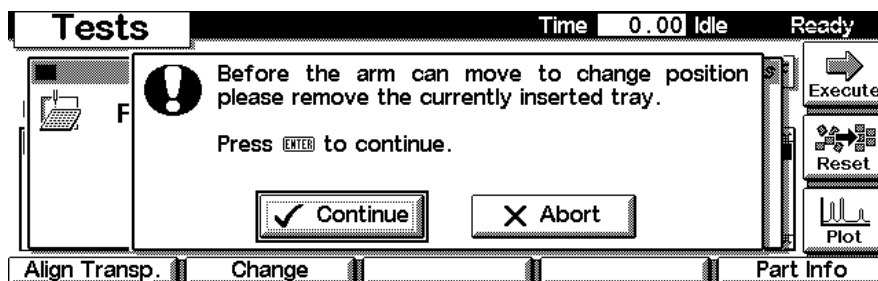
Use the **F1**-key (**Align Trans**) to perform an automatic alignment of the micro collector/spotter transport unit.

WARNING

An empty 4 well plate tray (G1364-84511 or G1364-84521) and the well plate adapter must be installed, when performing the alignment. The well plates must be removed from the micro collector/spotter tray before performing the transport unit alignment. If the well plates are installed during the alignment procedure, the transport unit might be damaged.

The alignment process may take several minutes. Changes must be acknowledged with **F6 (Done)**.

To exchange the capillary, the well plate adapter or the capillary carrier assembly the arm can be moved to the (**F2**) **Change** position, where the user has access to these parts. The tray has to be removed.



Records) Use the **F1**-key (**EMF**) to set EMF limits. Changes must be acknowledged with **F6 (Done)**.

3 Control Module Screens of the Fraction Collector

Screens available from the System view

Records

Time0.00IdleReady

Module	Product#	Serial#	Version	On-Time
Controller	G1323B	CN23605001	B.04.02	0d 00:14h
AF Collector	G1364D*	DE43640110	A.06.01 [012]	0d 00:14h

Records Time 0.00 Idle Ready

Automated Fraction Collector Error Logbook

Message	Date	Time
Initialization failed	10/21/05	16:38:23
Touchpad malfunction	10/21/05	16:38:21
Thermostat contact lost	10/21/05	16:32:04
Thermostat contact lost	10/21/05	16:32:03
Thermostat contact lost	10/21/05	16:32:02

Print

Records		Time	0.00	Idle	Ready
<i>Automated Fraction Collector Maintenance Logbook</i>					
Message	Date	Time			
capillary G1315-68708 replaced.cl	05/23/05	15:16:49			
cover replaced on 10 may 05,AT	05/10/05	13:01:41			
*** end of logbook ***					

Add Print

3 Control Module Screens of the Fraction Collector

Screens available from the System view



4 Troubleshooting and Test Functions

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Status Indicators

Two status indicators are located at the front panel of the micro collector/spotter. The lower left indicates the power supply status, the upper right indicates the micro collector/spotter status.

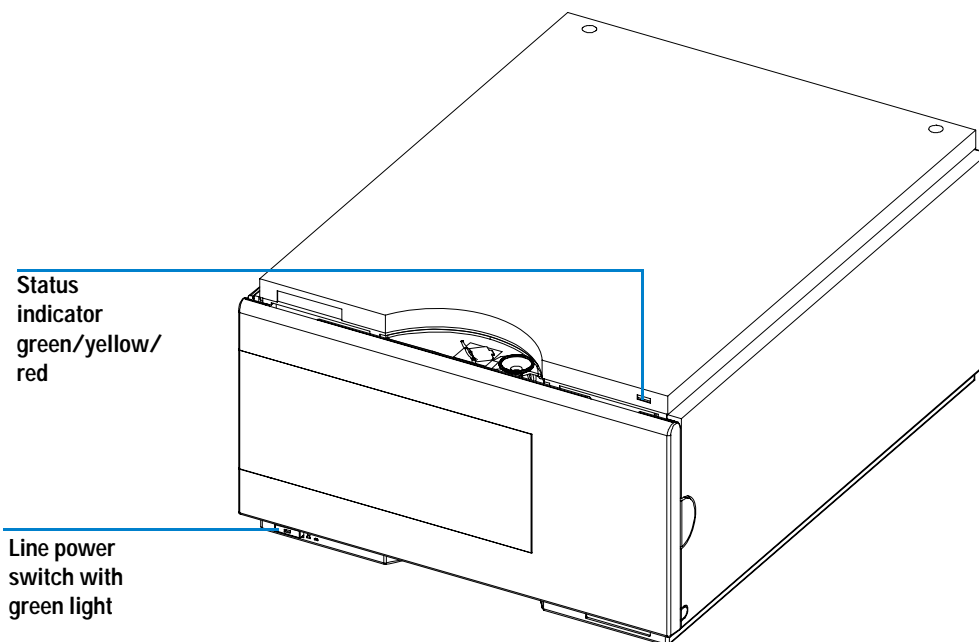


Figure 16 Location of Status Indicators

Power Supply Indicator

The power supply indicator is integrated into the main power switch. When the indicator is illuminated (*green*) the power is ON.

Instrument Status Indicator

The instrument status indicator indicates the current instrument condition:

- When the status indicator is OFF (and power switch light is on), the instrument is in a *prerun* condition, and is ready to begin an analysis.
- A *green* status indicator indicates the instrument is performing an analysis (*run* mode).
- A *yellow* status indicator indicates a *not-ready* condition. The instrument is in a not-ready state when it is waiting for a specific condition to be reached or completed (for example, front door not closed), or while a self-test procedure is running. The next programmed injection will be inhibited until the not-ready condition has been cleared.
- An *error* condition is indicated when the status indicator is *red*. An error condition indicates the instrument has detected an internal problem which affects correct operation of the instrument. Usually, an error condition requires attention (for example, leak, defective internal components). An error condition always interrupts the analysis and prevents the next run after the current run is finished.
- A *flashing yellow* status indicator indicates that the module is in the *resident mode*. Call your local service provider for assistance upon observing this error condition.
- A *flashing red* status indicator indicates an during the *startup* procedure of the module. Call your local service provider for assistance upon observing this error condition.

Deactivating the door lock sensor

The Agilent 1200 Series micro collector/spotter G1364D allows the user to operate the instrument with an open front door. In order to use this option the door lock sensor has to be disabled. Type the following command into the ChemStation Command Line

doorunlock

The door lock sensor should only be disabled for special applications. It should be activated again, if this operation mode is no longer necessary. To activate the door lock sensor again, type

doorlock

If the door lock sensor is disabled, the instrument will stay in the READY status while the door is open. However the fraction collector will always startup in the default mode (door lock on) after it has been power cycled.

WARNING

If the door lock sensor is disabled, the fraction collector arm will move while the front door is open. To avoid personal injury, keep fingers away from the needle area during fraction collector operation.

All users must be informed about this potential risk.

Maintenance Functions

Some maintenance procedures require the needle arm and needle carrier to move to specific positions to enable easy access to these components. The maintenance functions move these assemblies into the appropriate maintenance position.

NOTE

All trays have to be removed before starting the Maintenance dialog.

In the ChemStation the micro collector/spotter maintenance positions can be selected from the **Maintenance** menu in the **Diagnosis** view (see [Figure 17](#)). In the Control Module the functions can be selected in the **Test** screens of the micro collector/spotter.

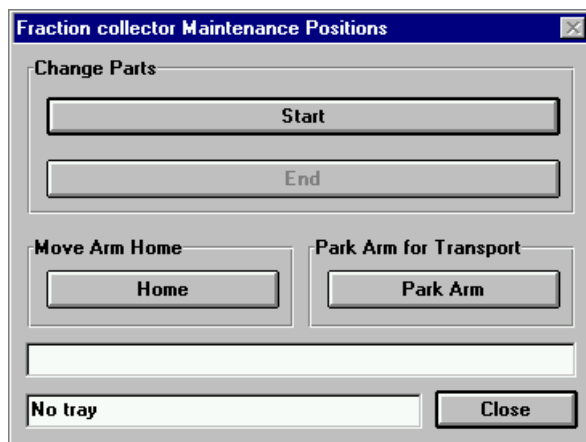


Figure 17 Fraction Collector Maintenance Positions dialog box

Change Parts.

If you click **Start** the transport unit will move upwards, the needle carrier assembly will move to the front center and then turn off the rotary motor to allow free rotation of the arm. This position enables easy access...

- to change the capillary guiding assembly
- to change the micro collector/spotter capillary
- to change the capillary carrier assembly
- to change the micro collector/spotter lamp assembly
- to replace the well plate adapter with the spotting adapter or vice versa
- to align the capillary length and install the spotting tip.

After the maintenance or repair task has been finished, click **End** to reset the micro collector spotter and move the arm to the waste port.

Home Position

This maintenance function moves the arm up and to the right rear for better access and exchange of the trays.

Park Arm

This maintenance position moves the arm to the park position at the upper rear left side of the tray for transporting or shipping the micro collector/spotter.

Transport Unit Self Alignment

The transport unit alignment is required to compensate for larger deviations in positioning the needle carrier assembly. This might be necessary after disassembling the system or when you exchange the transport unit, the needle carrier assembly or the MTP main board.

This function is in the diagnose screen of the ChemStation or the Control Module.

WARNING

The sample transport self alignment requires the 4-well-plate tray (G1364-84511 or G1364-84521). All well plates **MUST** be removed!

If the Transport Unit Self Alignment is started with well plates on the tray, the alignment procedure is aborted **WITHOUT** error message.

When is a Transport Unit Self Alignment Necessary?

The sample transport self alignment is required after disassembling the module or when you exchange:

- The transport unit.
- The needle/capillary carrier assembly.
- The MTP main board.

How to perform a Transport Unit Self Alignment?

Steps	Comments
1 If the transport unit has been exchanged or if it is strongly misaligned, set the 8-bit configuration switch to the Forced Cold Start Configuration.	See Service Manual G1364-90110 for details.
2 Install the 4-well-plate tray (G1364-84511 or G1364-84521)	IMPORTANT: Remove all plates!
3 Ensure that the wellplate adapter is correctly assembled	

How to perform a Transport Unit Self Alignment?

Steps	Comments
4 Select the Maintenance menu in the Diagnosis view of the Agilent ChemStation.	
5 In the menu choose Fraction Collector > Transport Alignment... to start the automated procedure.	The Transport Alignment Procedure takes approximately 10-15 minutes
6 Set the 8-bit configuration switch to the default setting.	See Service Manual G1364-90110 for details.

Arm Length Calibration

The Arm length calibration is only be required for 384 well plates. This procedure calibrates for the arm length of the capillary carrier assembly. The arm length calibration is required...

- after disassembling the transport unit, capillary carrier assembly.
- if problems with the positioning of the capillary tip occur - in particular on plate 4 (back right).

We recommend to switch on the lamp during the calibration. Open the command line in the ChemStation and type

`microafclampon` to switch the lamp ON
`microafclamppoff` to switch the lamp OFF

WARNING

During the arm length calibration the door lock sensor is disabled. The fraction collector arm will move while the front door is open. In addition the side panel can be removed during this procedure. To avoid personal injury, keep fingers away from the needle area during arm length calibration.

All users must be informed about this potential risk.

To perform an arm length calibration for 384-well plates

Step	Note
1	Before starting the arm length calibration procedure put two 384 well plates into the front positions (1 and 3) of the well plate tray. Select the type of these plates in the fraction collector configuration dialog.
2	Switch to the Diagnosis View of the Agilent ChemStation Software.

4 Troubleshooting and Test Functions

Arm Length Calibration

To perform an arm length calibration for 384-well plates (continued)

Step	Note
3 Select Tests... from the Diagnosis menu.	
4 In the Test Selection dialog box select Micro AFC - G1364D from the drop-down list.	
5 Select arm length calibration (384 well plates) and click on Start .	
6 Start the arm length calibration and follow the instructions on the screen.	Click on Explain to get detailed instructions and information
7 Click the Plate 1 button in the Arm Length Calibration dialog box (Figure 18 on page 99)	The capillary carrier will move to well position A7 on plate 1
8 Use the left and right button in the dialog box (Figure 18) to adjust the capillary tip to the center of the well	
9 Click the Plate 3 button in the Arm Length Calibration dialog box (Figure 18 on page 99). in the Arm Length Calibration dialog box (Figure 18 on page 99)	The capillary carrier will move to well position A18 on plate 3
10 Use the left and right button to adjust the capillary tip to the center of the well.	
11 After adjusting the capillary this way the determined values are stored by pressing the OK button.	It might take some time to store the positions on the micro collector/spotter.

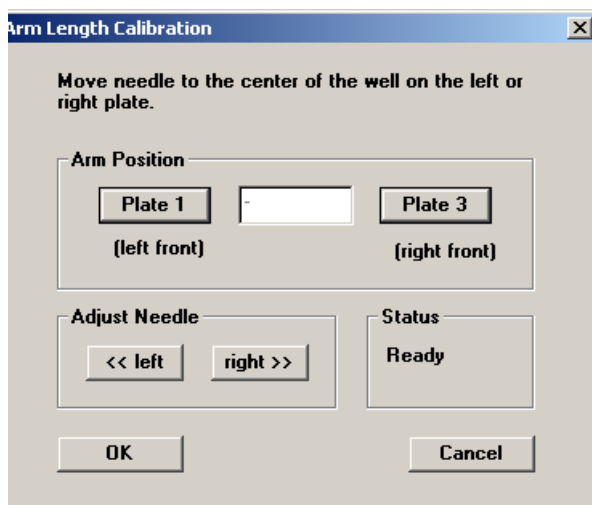


Figure 18 Arm Length Calibration Interface

Position Accuracy Calibration for 384-Well Plates

We recommend to perform the Position Accuracy Calibration for 384-well plates...

- if the positioning is inaccurate
- after mayor repairs (e.g. exchange of transport assembly, capillary carrier or mainboard)
- after a firmware update.
- if a different type of 384-well plate is used.

We recommend switching on the lamp during the calibration. Open the command line in the ChemStation and type

microafclampon to switch the lamp ON
microafclampoff to switch the lamp OFF

WARNING

During the position accuracy calibration the door lock sensor is disabled. The fraction collector arm will move while the front door is open. In addition the side panel can be removed during this procedure. To avoid personal injury, keep fingers away from the needle area during position accuracy calibration.

All users must be informed about this potential risk.

To perform a position accuracy calibration for well plates :

Step	Note
1	Switch to the Diagnosis View of the Agilent ChemStation Software
2	Select Tests... from the Diagnosis menu
3	In the Test Selection dialog box select Micro AFC - G1364D from the drop-down list.

To perform a position accuracy calibration for well plates (continued):

Step	Note
4	Select position accuracy calibration (well plates) and click on Start .
5	Start the position accuracy calibration and follow the instructions on the screen. Click on Explain to get detailed instructions and information
6	Align the capillary tip to the center of each corner well with the corresponding positioning buttons. See Figure 19 on page 101. The height calibration is not required for well plates and therefore greyed out.
7	Repeat Step 6 for all configured plates

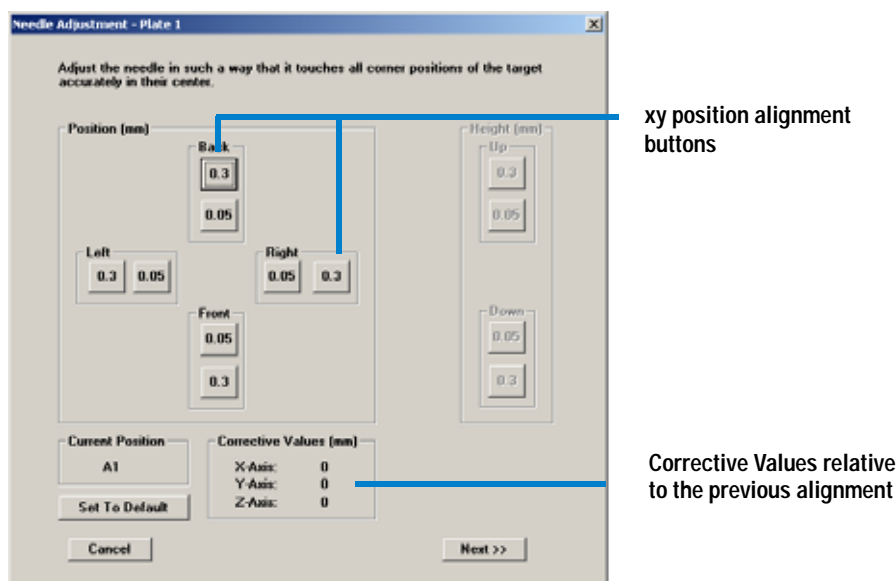


Figure 19 Position Accuracy Calibration Interface for well plates

Position Accuracy Calibration for MALDI Targets

The Position Accuracy Calibration for MALDI targets has to be performed, if

- a different type of MALDI Target is used.
- the positioning is inaccurate.
- after mayor repairs (e.g. exchange of transport assembly, capillary carrier or mainboard)
- after a firmware update.

We recommend to switch on the lamp during the calibration. Open the command line in the ChemStation and type

microafclampon to switch the lamp ON

microafclamppoff to switch the lamp OFF

WARNING

During the position accuracy calibration the door lock sensor is disabled. The fraction collector arm will move while the front door is open. To avoid personal injury, keep fingers away from the needle area during position accuracy calibration.

All users must be informed about this potential risk.

CAUTION

The position accuracy depends on the temperature. We recommend to perform the position accuracy alignment at the same temperature as the operation of the instrument. If a fraction collector thermostat is present, it should be switched on 30 minutes before the calibration to ensure constant temperature conditions in the module.

Table 8 To perform a position accuracy calibration for MALDI targets

Step	Note
1	Switch to the Diagnosis View of the Agilent ChemStation Software
2	Select Tests... from the Diagnosis menu

Table 8 To perform a position accuracy calibration for MALDI targets (continued)

Step	Note
3 In the Test Selection dialog box select Micro AFC - G1364D from the drop-down list.	
4 Select position accuracy calibration (MALDI) and click on Start .	
5 Start the position accuracy calibration and follow the instructions on the screen.	Click on Explain to get detailed instructions and information
6 Align the xy position of the capillary tip to the center of each corner position of the MALDI Target or to each pinhole on the MALDI calibration plates with the corresponding positioning buttons.	See Figure 20 on page 104. <ul style="list-style-type: none"> • for Agilent Technologies AP-MALDI targets use calibration plate 5023-0214 • for Micromass MALDI targets use calibration plate 5023-0215 • for Bruker MALDI Targets use calibration plate 5023-0208 • for Applied Biosystems MALDI targets use calibration plates 5023-0209 or 5023-0213 • If a calibration plate is not available use the MALDI Target and align the capillary to the center of each corner position.
7 Align the height (z position) of the capillary tip.	The capillary tip should be aligned just above the MALDI Target leaving enough space for sheet of paper to fit between the capillary tip and the target.
8 Repeat Step 6-7 for all configured plates	

4 Troubleshooting and Test Functions

Position Accuracy Calibration for MALDI Targets

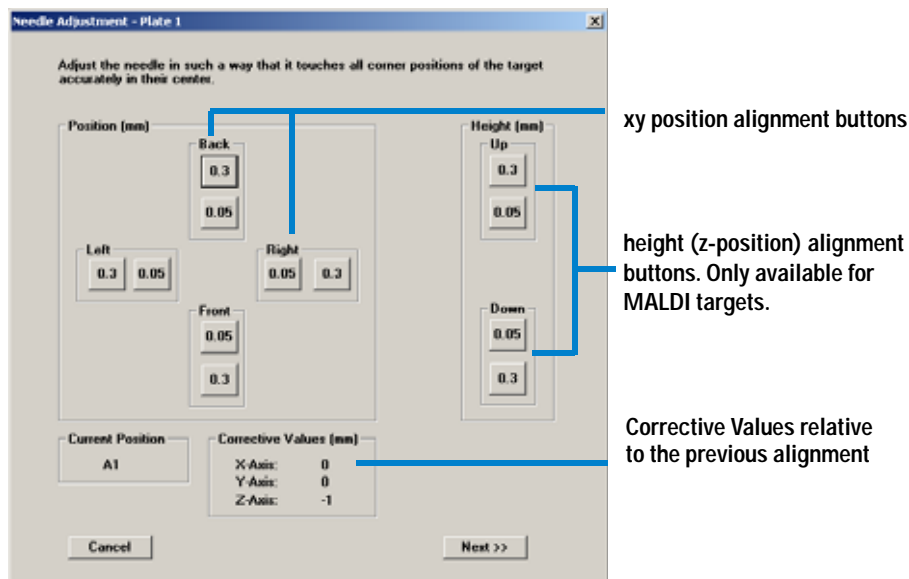


Figure 20 Position Accuracy Calibration Interface for MALDI Targets

Single Step Commands

Some movements of the fraction collection or spotting sequence can be done under manual control. This is useful during troubleshooting where close observation of each of the fraction collection steps is required to confirm a specific failure mode or verify successful completion of a repair.

Each step command actually consists of a series of individual commands which move the micro collector/spotter components to predefined positions enabling the specific step to be done.

In the ChemStation the step commands can be selected from the “**Test Selection Box**” (see [Figure 21](#)) in the **Diagnosis** display. In the Control Module the step commands can be accessed from the pull-down menu in the micro collector/spotter “**Test**”.

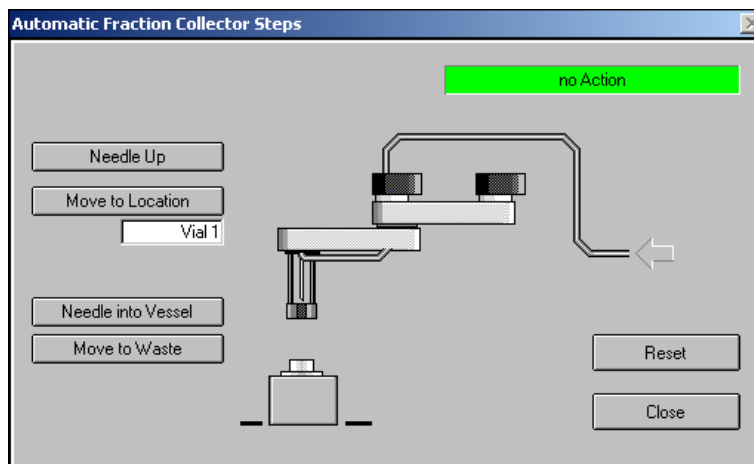


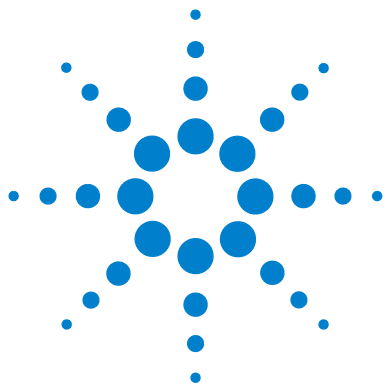
Figure 21 Fraction Collector Step Commands

4 Troubleshooting and Test Functions

Single Step Commands

Table 9 Step Commands

Step	Action	Comments
Needle Up	Lifts the needle arm to the upper position.	
Needle into vessel	Lowers the needle into the specified vessel.	
Needle to rinse / flush port	Moves the needle to the rinse / flush port.	



5 Maintenance and Simple Repairs

Introduction into Repairing the Micro Collector/Spotter	108
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Introduction into Repairing the Micro Collector/Spotter

Simple Repairs

The micro collector/spotter is designed for easy repair. The most frequent repairs and maintenance tasks such as changing the capillary, the flap septum or the waste tubing can be done from the front of the instrument with the instrument in place in the system stack. These repairs are described in [“Simple Repairs Procedures”](#) on page 116.

WARNING

When opening capillary or tube fittings solvents may leak out. Please observe appropriate safety procedures (for example, goggles, safety gloves and protective clothing) as described in the material handling and safety data sheet supplied by the solvent vendor, especially when toxic or hazardous solvents are used.

WARNING

Regularly inspect the capillary and exchange it if they is worn out or shows visible signs of damage.

WARNING

To prevent personal injury, the power cable must be removed from the instrument before removing the micro collector/spotter cover. Do not connect the power cable to the micro collector/spotter while the cover is removed.

Cleaning the Micro Collector/Spotter

The micro collector/spotter covers should be kept clean. Cleaning should be done with a soft cloth slightly dampened with water or a solution of water and a mild detergent. Do not use an excessively damp cloth from which liquid could drip into the micro collector/spotter.

WARNING

Do not let liquid drip into the micro collector/spotter. It could cause a shock hazard or damage to the micro collector/spotter.

Cleaning the waste port

Frequency	The frequency depends strongly on the application.
Tools required	None
Parts Required	None

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation.

CAUTION

If solvents with high concentrations of matrix, buffer or salt are used, the capillary has to be flushed thoroughly with salt free water after analysis. Such a procedure prevents the capillary and the waste port from clogging.

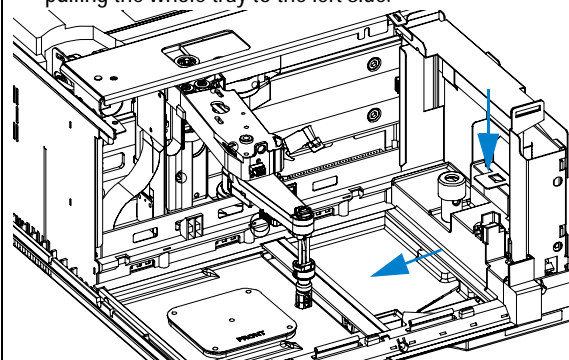
NOTE

For low flow rates and for MALDI spotting with online matrix delivery, we recommend to use the plug to close the waste container instead the waste tubing. The plug (G1364-26105) is included in the micro collector/spotter accessory kit.

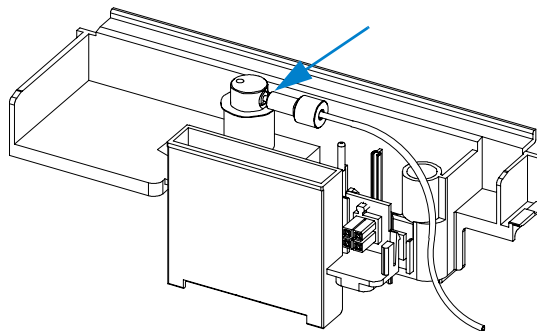
5 Maintenance and Simple Repairs

Introduction into Repairing the Micro Collector/Spotter

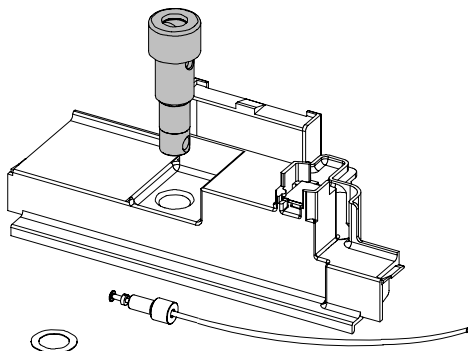
1 The internal tray is located on the right side of the fraction collector. It can easily be removed by lifting the clip and pulling the whole tray to the left side.



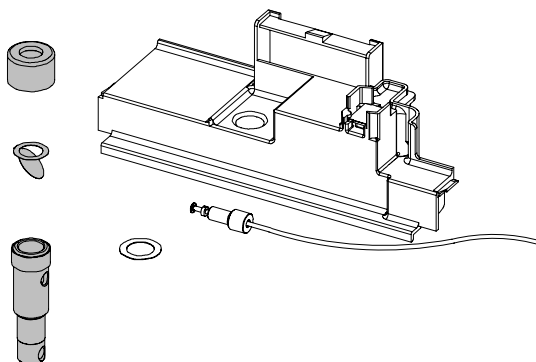
2 Turn the tray upside down, unscrew the tubing and remove the washer.



3 Now the whole waste vial can be removed as well.



4 After unscrewing the cap the waste port can be further disassembled.



To clean the wash port and the waste tubing

- For a thorough cleaning of all components they can be sonicated for 15 minutes in an ultrasonic bath filled with water.
- Another way to get rid of plugging, is to flush the waste capillary from the exit with a water filled syringe. Attach the syringe at the end of the waste tubing using a 1.6 mm OD capillary and flush the waste tubing and the waste port thoroughly.

In order to avoid flooding of the fraction collector a tissue should be placed besides the internal tray close to the spill.

On completion of this procedure:

- Assemble the cleaned waste tubing
- Install the internal tray in the fraction collector as shown in [step 4](#) on [page 125](#)
- **IMPORTANT: Make sure that the waste tubing is guided correctly through the waste tubing channel to the waste tubing outlet as shown in [step 4](#) and [step 5](#) on [page 125](#).**

Exchanging the micro collector/spotter lamp assembly

Frequency	When lamp is defective or not required in the micro collector/spotter
Tools required	None
Parts Required	Micro collector/spotter lamp assembly G1364-60014

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation.

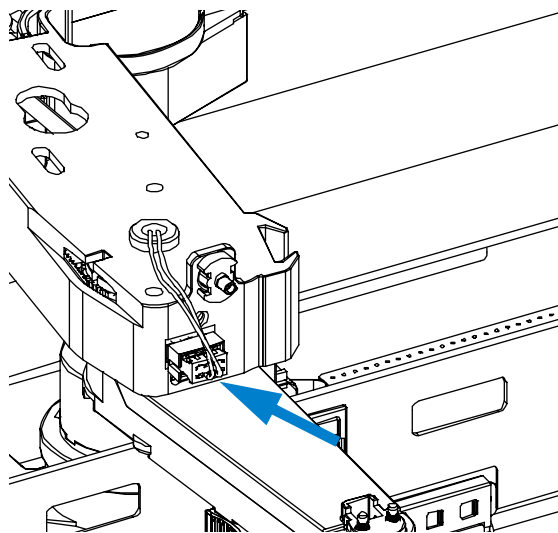
WARNING

Explicitly follow the described installation procedures to maximize the lifetime of the fraction collector capillary and to avoid potential spills or fraction losses. Regularly inspect the capillary and exchange it if it is worn out or shows visible signs of damage.

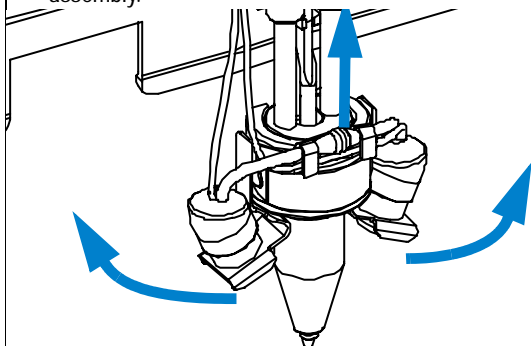
Before beginning this procedure:

- Position the transport unit of the micro collector/spotter in the **“Home Position”** (see **“Maintenance Functions”** on page 93).
- Remove all installed trays.
- Position the transport unit of the micro collector/spotter in the **“Change Parts Position”** (see **“Maintenance Functions”** on page 93).
- Turn OFF the instrument.
- Unscrew the micro collector/spotter capillary from the flow cell of the detector.
- Remove the fraction collector/spotter capillary as described in **“Replacing Micro Collector/Spotter Capillary Assembly”** on page 117

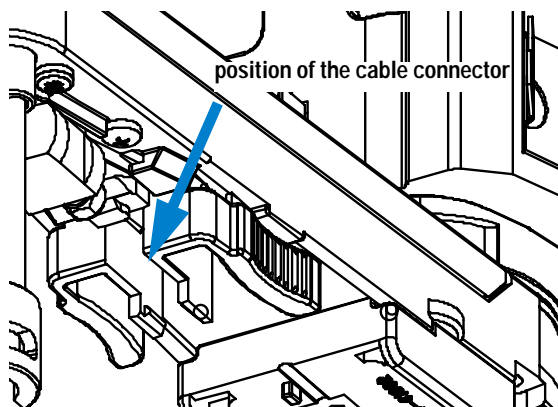
- 1** Unplug the lamp power cable from the connector at the front of the z-arm



- 2** To disassemble the lamp carefully bend up the two metal sheets, that hold the LEDs. Then push up the lamp assembly.



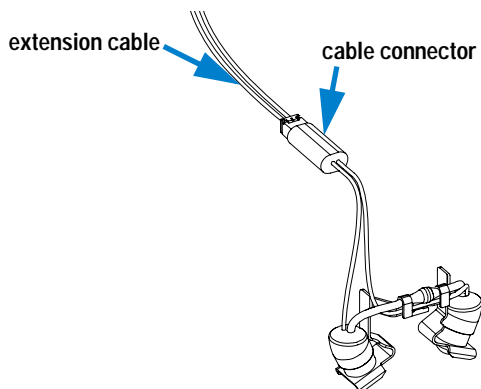
- 3** Un-clip the cable connector which is located between the plastic clips of the capillary guiding assembly.



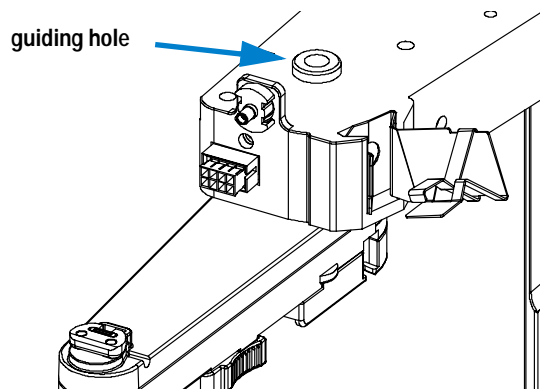
5 Maintenance and Simple Repairs

Exchanging the micro collector/spotter lamp assembly

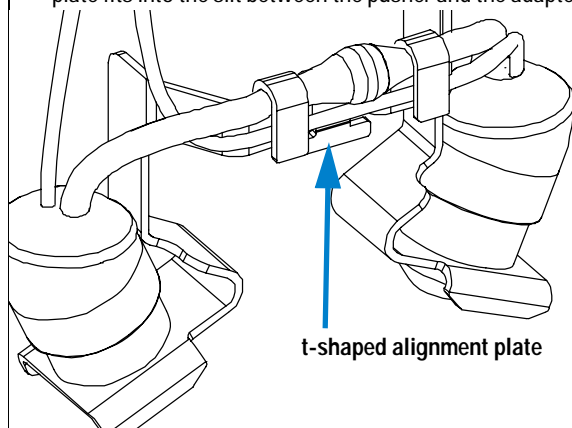
4 Disconnect the lamp assembly from the extension cable at the cable connector and pull the cable through the guiding hole in the carrier assembly.



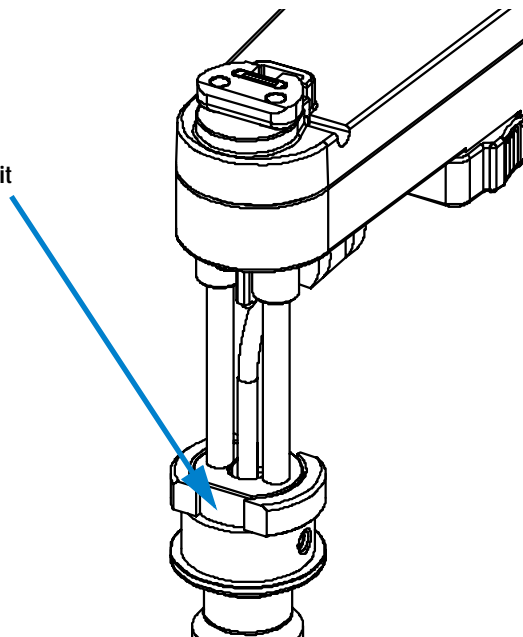
5 To install the new lamp assembly unplug the connector as described in [step 4](#) and slide the cable from the top through the hole in the carrier assembly.



6 Carefully bend up the two metal sheets of the lamp assembly, that hold the LEDs. Then slide the assembly onto the spotting adapter, that the t-shaped alignment plate fits into the slit between the pusher and the adapter.



slit



- Connect the lamp assembly to the extension cable.
- Push the cable connector in-between the yellow plastic clip as described in [step 3](#) of this procedure.
- Plug the lamp power cable into the connector at the front of the z-arm.
- Install the fraction collector/ spotter capillary as described in [“Replacing Micro Collector/Spotter Capillary Assembly”](#) on page 117
- To activate the lamp switch open the command line in the ChemStation and type
microafclampon
to switch the lamp ON
microafclampoff
to switch the lamp OFF

Simple Repairs Procedures

The procedures described in this section can be done with the micro collector/spotter in place in the stack.

Table 10 Simple Repair Procedures

Procedure	Typical Frequency	Notes
Replacing the fraction collector capillary	When worn out, blocked or damaged. When using a capillary with different diameter.	See “Replacing Micro Collector/Spotter Capillary Assembly” on page 117
Exchanging the capillary guiding assembly	When bent or damaged.	See “Exchanging the Capillary Guiding Assembly” on page 120
Exchanging the internal tray	When flow delay sensor defective	See “Exchanging the Internal Tray” on page 124
Exchanging the flap septum and the waste tubing	When defective or contaminated	See “Exchanging the Flap Septum and the Waste Tubing” on page 122

R

Replacing Micro Collector/Spotter Capillary Assembly

Frequency	When contaminated, blocked, worn out or visibly damaged. When using a new capillary with different diameter.
Tools required	None
Parts Required	Capillary Assembly 25 μ m ID (G1364-87304), Capillary Assembly 50 μ m ID (G1364-87305) or Capillary Assembly 100 μ m ID (G1364-87306)

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation.

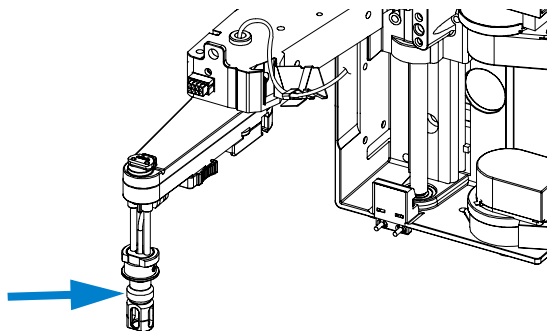
WARNING

Thoroughly follow the described installation procedures to maximize the lifetime of the fraction collector capillary and to avoid potential spills or fraction losses. Regularly inspect the capillary and exchange it if it is worn out or shows visible signs of damage.

Before beginning this procedure:

- Position the transport unit of the micro collector/spotter in the “Home Position” (see “Maintenance Functions” on page 93).
- Remove all installed trays from the tray base.
- Position the transport unit of the micro collector/spotter in the “Change Parts Position” (see “Maintenance Functions” on page 93).
- Turn OFF the instrument.
- Unscrew the micro collector/spotter capillary from the flow cell of the detector.

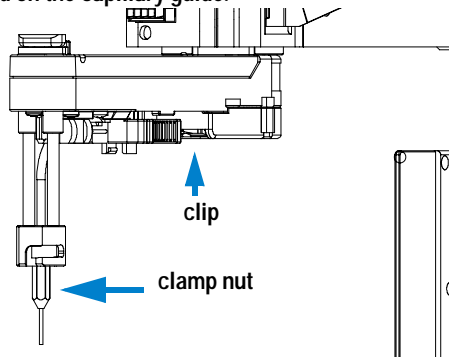
- 1 Remove the wellplate adapter from the Capillary Carrier Assembly.



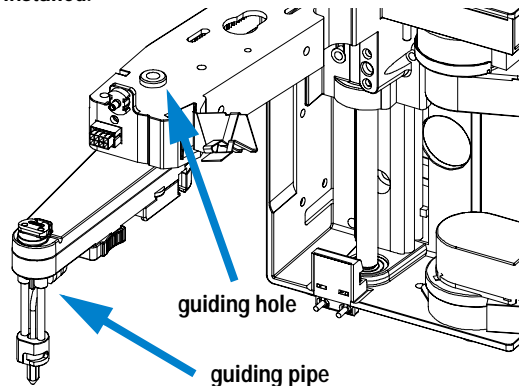
5 Maintenance and Simple Repairs

Simple Repairs Procedures

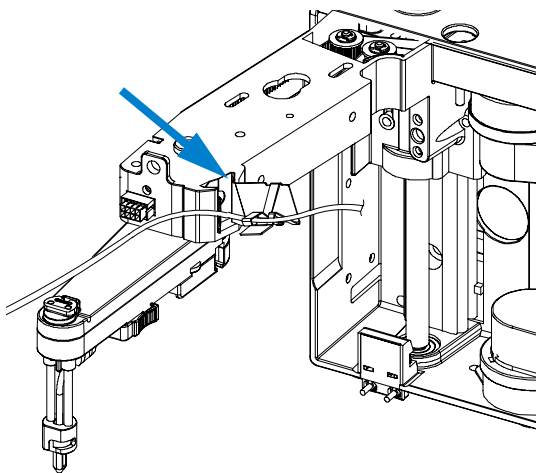
- 2 Remove the clamp nut and remove the capillary from the clip at the bottom of the carrier assembly.
Do not adjust the capillary while the clamp nut is tightly mounted on the capillary guide.



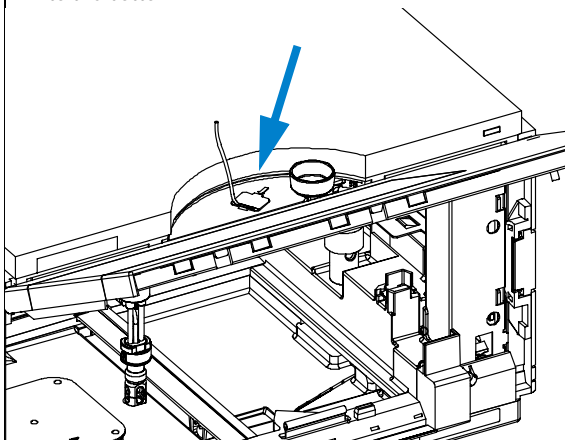
- 3 Pull the capillary out off the capillary guiding pipe and out off the guiding hole in z-arm of the transport assembly.
Do not tighten the clamp nut without a capillary installed.



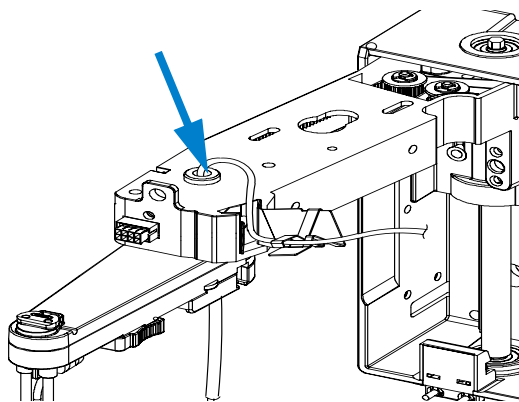
- 4 Press the metal sheets of fixation spring together and push the capillary assembly out of the holder at the z-arm of the transport assembly.



- 5 Unplug the capillary assembly from the bolt carrier (located on the top center of the front cover) by moving the snapper as indicated by the arrows. Slide the Capillary out to the bottom.

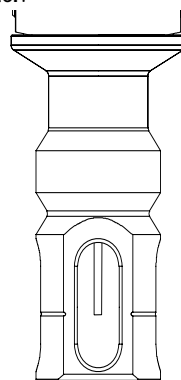


6 Push the new capillary through the bolt carrier as shown in [step 5](#) of this procedure. Then attach the fixation spring at the z-arm of the transport assembly and push the capillary through the guiding hole.

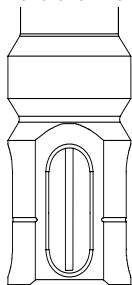


7 Push the capillary through the guiding fitting assembly and attach the wellplate adapter. Then adjust the position of the capillary tip.

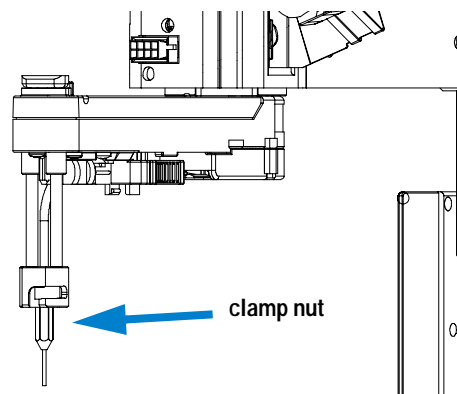
For highest accuracy that is required for 384 well plates the tip must be close to the groove in the middle of the wellplate adapter.



8 If you are using a deep well plate, you should push the capillary through the guiding assembly, that the tip is close to the bottom of the wellplate adapter frame. The tip must stay within the oval frame.



9 If the capillary tip is at the correct position, remove the wellplate adapter, tighten the clamp nut and remount the adapter.



Exchanging the Capillary Guiding Assembly

Frequency	When Capillary guiding assembly is bent or damaged
Tools required	None
Parts required	Capillary guiding assembly, G1364-87303

WARNING

To avoid personal injury, keep fingers away from the needle area during micro collector/spotter operation.

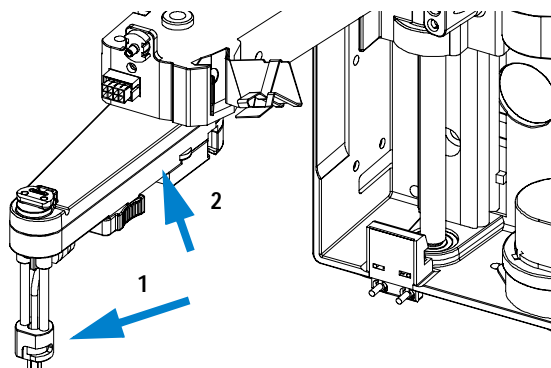
CAUTION

Regularly inspect the Fraction Collector Capillary and exchange it if it is worn out or shows visible signs of damage.

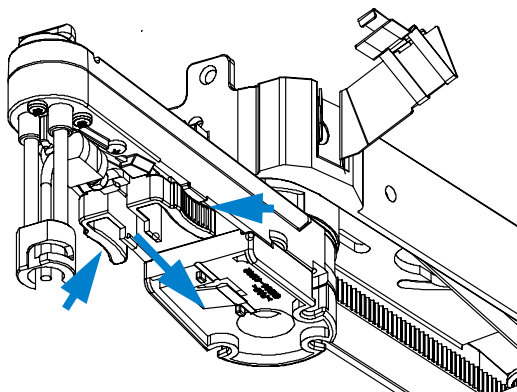
Before beginning this procedure:

- Move the transport unit of the micro collector/spotter in the “Home Position” (see “Maintenance Functions” on page 93).
- Remove all installed trays from the tray base.
- Position the transport unit of the micro collector/spotter in the “Exchange Parts Position” (see “Maintenance Functions” on page 93).
- Turn OFF the instrument.
- **IMPORTANT: Do not adjust the capillary while the clamp nut is tightly mounted on the capillary guide**

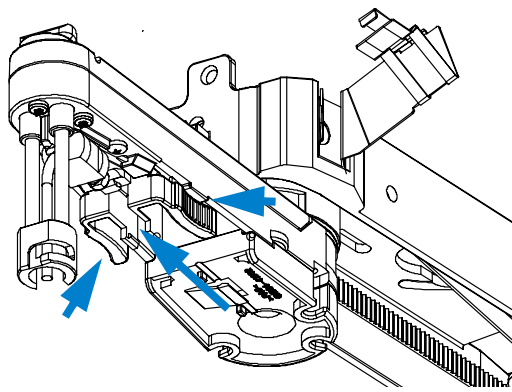
- 1 Remove the wellplate adapter and the clamp nut. Then pull the capillary from the clip at the bottom of the carrier assembly as well as the capillary guiding assembly.



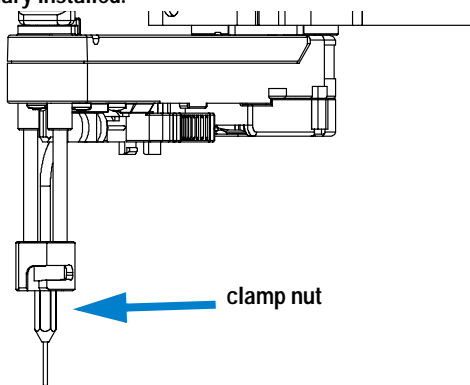
2 Press the yellow clips of the capillary guiding assembly together and pull the assembly towards the rear of the capillary carrier assembly.



3 Slide the new capillary guiding assembly into the holder of the capillary carrier assembly. Make sure to push it all the way to the front.



4 Reinstall the Capillary, the clamp nut and the wellplate adapter.
IMPORTANT: Do not tighten the clamp nut without a capillary installed.



On completion of this procedure:

- Re-install the tray(s) in the tray base.
- Start the instrument.
- Close the front cover.

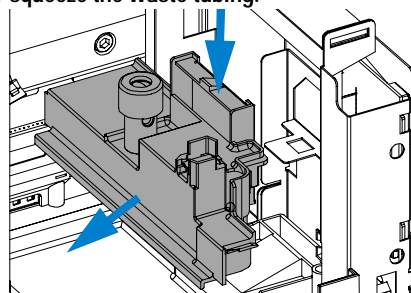
Exchanging the Flap Septum and the Waste Tubing

Frequency	When defective or contaminated When waste port is clogged
Tools required	None
Parts required	Flap Septum (G1364-27107) Waste Tubing (G1364-86711)

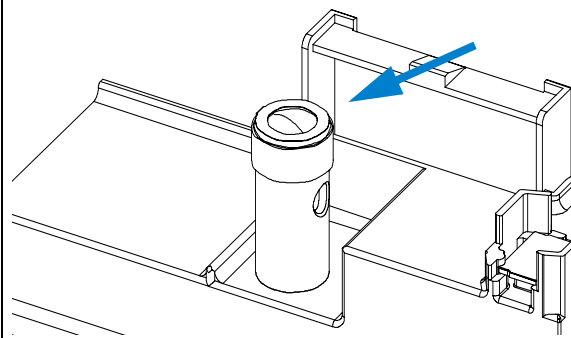
Before beginning this procedure:

- Position the transport unit of the micro collector/spotter in the “Home Position” (see “Deactivating the door lock sensor” on page 92) and remove all installed trays from the tray base.
- Turn OFF the instrument.

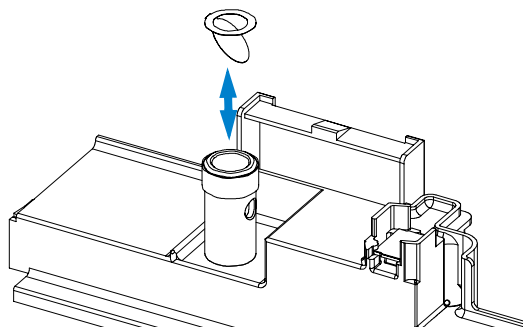
- 1 Remove the internal tray from the instrument, see “Exchanging the Internal Tray” on page 124. Don’t snap off or squeeze the waste tubing.



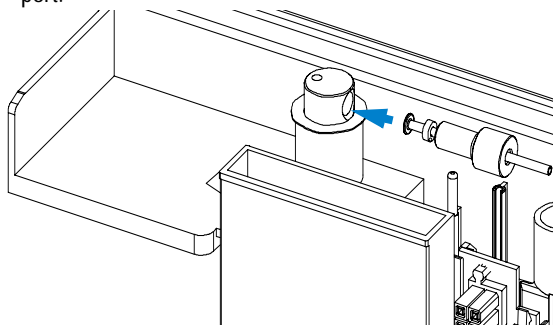
- 2 Remove the screw cap from the waste port



- 3 Exchange the Flap Septum, reassemble and tighten the screw cap



4 To exchange the waste tubing turn the internal tray upside down and unscrew the waste tube fitting from the waste port.



On completion of this procedure:

- Assemble the new waste tubing
- Install the internal tray in the fraction collector as shown in [step 4](#) on [page 125](#)
- **IMPORTANT:** Make sure that the waste tubing is guided correctly through the waste tubing channel to the waste tubing outlet as shown in [step 4](#) and [step 5](#) on [page 125](#).

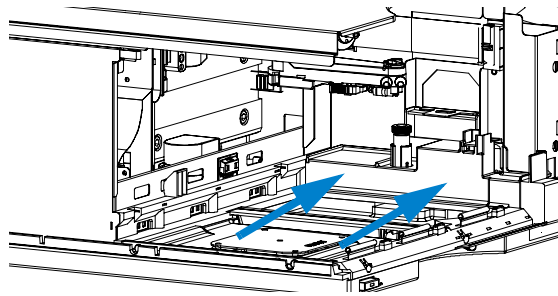
Exchanging the Internal Tray

Frequency	When defective
Tools required	None
Parts required	Internal Tray micro scale (G1364-63115)

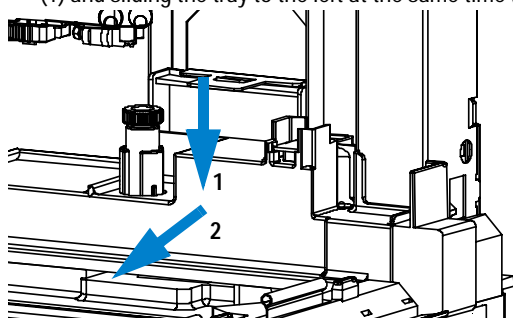
Before beginning this procedure:

- Position the transport unit of the micro collector/spotter in the “Home Position” (see “Maintenance Functions” on page 93).
- Remove all installed trays from the tray base.
- Turn OFF the instrument.

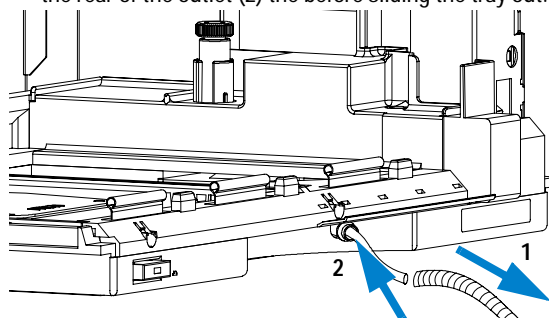
- 1 Locate the internal tray assembly with the rinse funnel and flow delay sensor in the bottom of the right front corner of the instrument.



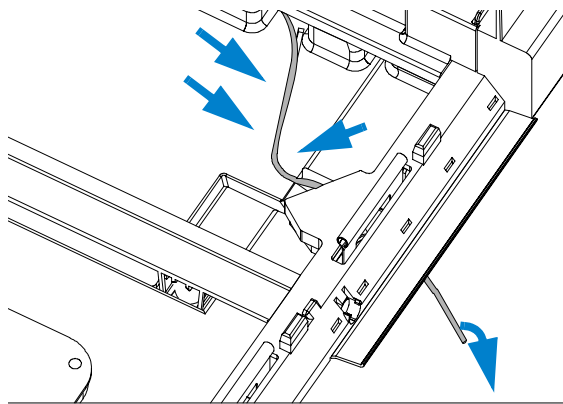
2 Remove the internal tray by pushing down the plastic holder that holds it in position underneath the metal latch (1) and sliding the tray to the left at the same time (2).



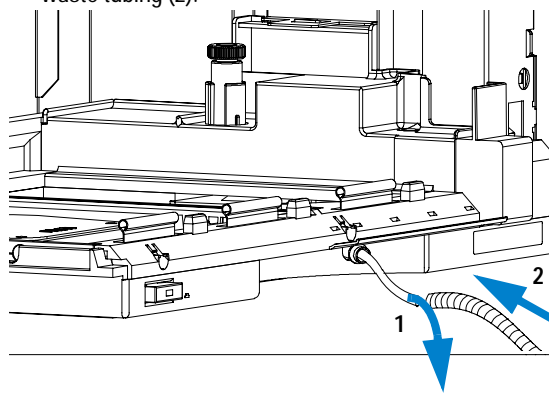
3 Remove the corrugated waste tubing from the front of the instrument (1) and slide the internal tray's waste tubing to the rear of the outlet (2) before sliding the tray out.



4 Install the new tray by sliding it into position underneath the metal latch that holds it. The waste tubing from the internal tray should be guided as shown.



5 Make sure that the waste tubing is slid all the way through the outlet (1). Its end should be over the edge and below the level of the laboratory desk that the system stands on to avoid any back flow of solvent. Re-install the corrugated waste tubing (2).

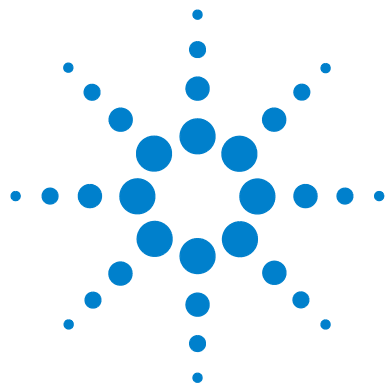


On completion of this procedure:

- Re-install the tray(s) in the tray base.
- Start the instrument.
- Close the front cover.

5 Maintenance and Simple Repairs

Simple Repairs Procedures



6 Parts and Materials

Supported Trays for the Micro Collector/Spotter [128](#)

List of Recommended Plates [130](#)

MALDI Spotting Accessories [131](#)

Supported MALDI Targets [133](#)

Transport Unit Assembly [135](#)

Internal Tray Assembly [136](#)

Micro Collector/Spotter Accessory Kit [137](#)



Supported Trays for the Micro Collector/Spotter

Table 11 Trays for the Micro Collector/Spotter

Item	Description	Part Number
1	Full tray for 4 well plates	G1364-84521
2	Full tray for 4 MALDI plate carriers, adjustable	G1364-84511
3	Std. tray for 2 well plates + 10 2ml vials	G1367-60011
4	Adapter air channel (installed underneath plug channel, if the fraction collector is used with the thermostat)	G1329-43200
5	Plug channel	G1364-47200

NOTE

Only one type of well-plate can be used at a time.

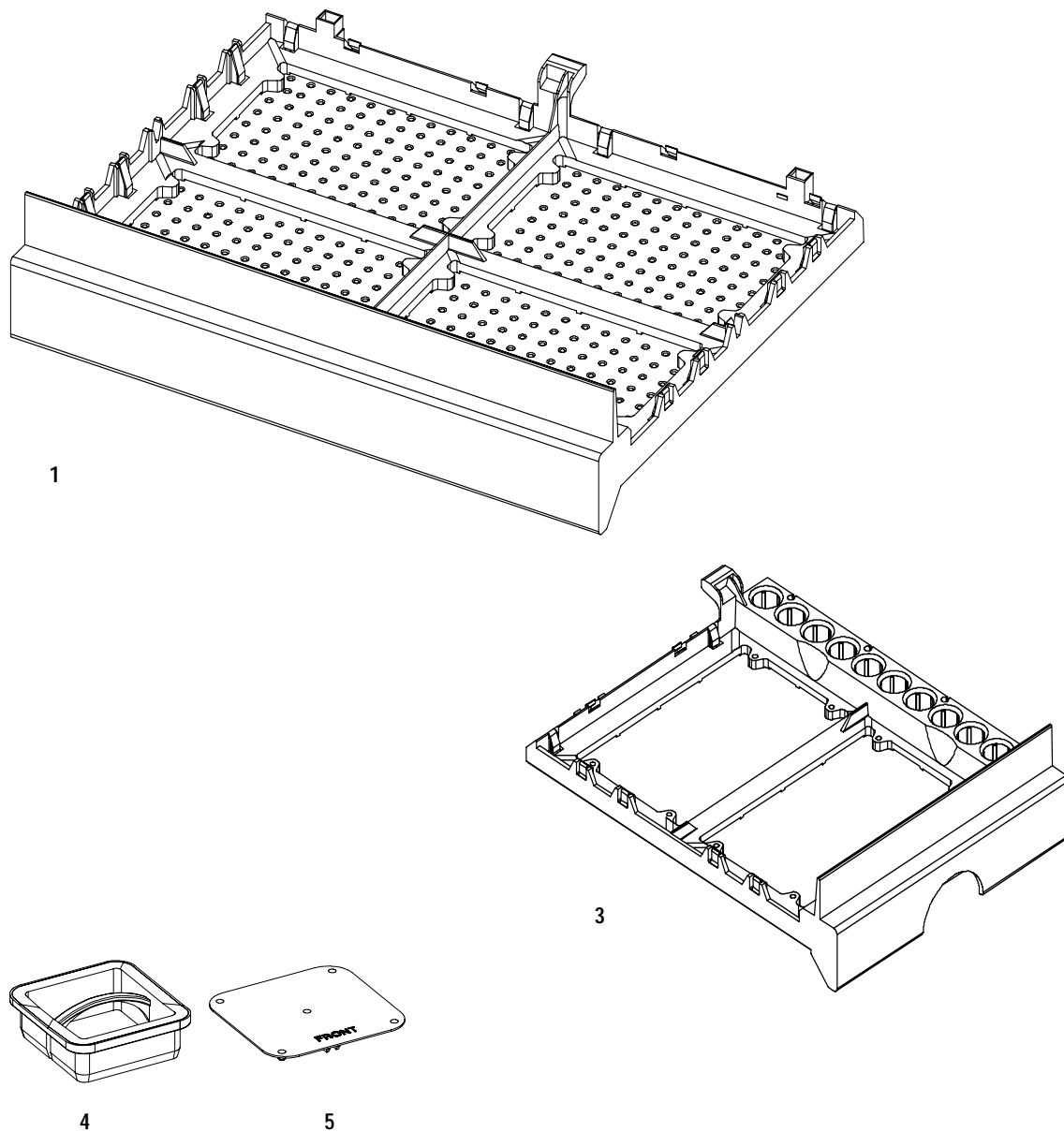


Figure 22 Trays

List of Recommended Plates

Table 12 Recommended Plates

Item	Description	Select in ChemStation	Volume (ml)	Package	Part Number
1	96 polypropylene well-plate	96Agilent	0.5	10	5042-1386
2	96 polypropylene well-plate	96Agilent	0.5	120	5042-1385
3	384 polypropylene well-plate	384Agilent	0.1	30	5042-1388
4	96 polypropylene conical-well plate	96EppendorfC	0.18	25	5042-8502
5	54 x 2ml vial plate	54VialPlate	1.5	6	G2255-68700

NOTE

Only one type of well-plates can be used at a time in one tray.

WARNING

If you are using flammable solvents, remove the plates from the micro collector/spotter after turning it OFF. You avoid the risk of building explosive gas mixtures in the instrument.

MALDI Spotting Accessories

Table 13 MALDI Plate Carriers

Plate Type	Part Number
MALDI Plate Carrier for Agilent Technologies AP-MALDI Targets	5022-6543
MALDI Plate Carrier for Bruker AnchorChip	5022-6541
MALDI Plate Carrier for Bruker Prespotted AnchorChip	5022-6546
MALDI Plate Carrier for Applied Biosystems Targets (standard format)	5022-6542
MALDI Plate Carrier for Applied Biosystems Opti-TOF (MTP format)	5023-0238
MALDI Plate Carrier for Micromass Targets	5022-6544

Table 14 Calibration Plates

Plate Type	Part Number
Calibration Plate for Agilent Technologies AP-MALDI Targets	5023-0214
Calibration Plate for Bruker Targets	5023-0208
Calibration Plate for Applied Biosystems Targets (10x10, 20x20)	5023-0213
Calibration Plate for Applied Biosystems Targets (192, 96x2)	5023-0209
Calibration Plate for Micromass Targets	5023-0215

Table 15 Online Matrix Kit G1364-68706

Item	Part Number
External Contact BCD Board	G1351-68701
Cable (BCD Board to Syringe Pump)	5181-1536
Micro Connector T-Type	5042-8519

Table 15 Online Matrix Kit G1364-68706 (continued)

Item	Part Number
Peek coated fused silica capillary (550mm length, ID 125µm)	G1375-87318
Adapter 10-32 to 1/4-28	5042-8517
Syringe 1ml	5181-1541
Union to Luer Fitting	5042-8518
Needle, LL, 22/51/3 (2/pk)	5183-4614

Supported MALDI Targets

Table 16 Supported Targets for MALDI Spotting

Note: MALDI target plates for Applied Biosystems, Bruker and Micromass are not supplied by Agilent Technologies.
Dimensions for other targets can be configured in the ChemStation Software.

- **Agilent Technologies**
 - 96 Agilent for AP MALDI (G1972-60025)
- **Applied Biosystems**
 - 80 Applied Biosystems
 - 96 Applied Biosystems
 - 100 Applied Biosystems
 - 96x2a Applied Biosystems
 - 96x2b Applied Biosystems
 - 192 Applied Biosystems
 - 400 Perseptive Biosystems
 - 144 Applied Biosystems
 - 192 Opti-TOF target
 - 384 Opti-TOF target (MPT format)
 - 96 Opti-TOF target (MPT format)
- **Bruker**
 - 384 AnchorChip Sample
 - 384 AnchorChip Lock
 - 1536 AnchorChip
 - 384 Prespotted AnchorChip
- **Micromass**

Table 16 Supported Targets for MALDI Spotting (continued)

Note: MALDI target plates for Applied Biosystems, Bruker and Micromass are not supplied by Agilent Technologies.

Dimensions for other targets can be configured in the ChemStation Software.

- 96 Micromass Sample
 - 96 Micromass Lock
 - 84 Micromass Sample
 - 84 Micromass Lock
 - 84 Micromass Calib
-

Transport Unit Assembly

Table 17 Transport Unit Assembly (Micro Scale)

Item	Description	Part Number
1	Transport unit assembly (micro scale), includes items 2 - 4.	G1364-60020
2	Capillary carrier assembly (micro scale)	G1364-60023
3	Capillary Guiding Assembly	G1364-87303
4	Wellplate adapter	G1364-23203
5a	Fraction Collector Capillary Assembly (25 µm ID)	G1364-87304
5b	Fraction Collector Capillary Assembly (50 µm ID)	G1364-87305
5c	Fraction Collector Capillary Assembly (100 µm ID)	G1364-87306

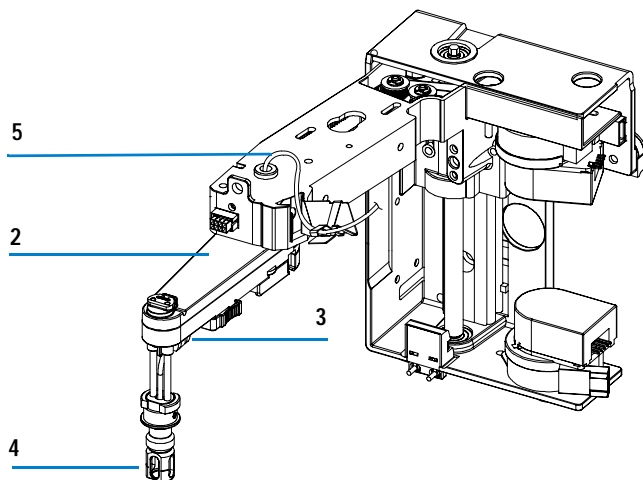


Figure 23 Transport Unit Assembly (Micro Scale)

Internal Tray Assembly

Table 18 Internal Tray Assembly (Micro Scale)

Item	Description	Part Number
1	Internal tray assembly (micro scale), includes items 2 – 6	G1364-63115
2	Waste tubing with fitting	G1364-86711
3	Washer	3050-2204
4	Screw cap	9301-1379
5	Flap septum	G1364-27107
6	Waste Container	G1364-22301

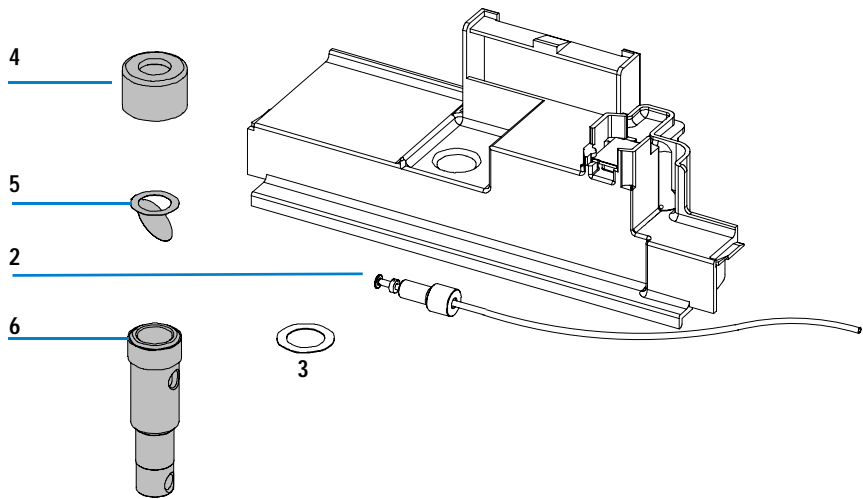


Figure 24 Internal Tray Assembly (Micro Scale)

Micro Collector/Spotter Accessory Kit

Table 19 Micro Collector/Spotter Accessory Kit Contents G1364-68715

Description	Quantity	Part Number
Hex key 2.0 mm	1	8710-2476
MALDI Spotting Adapter	1	G1364-83205
Flap Septum	1	G1364-27107
Waste Tubing 1.4 mm ID	1	G1364-86711
Waste tubing (1.2 m)*	1	5062-2463
CAN cable, 1 m	1	5181-1519
Sticking clamp for corrugated waste tubing (large)†	3	5065-9976
Sticking clamp for waste tubing (small)‡	3	5065-9976
ESD wrist strap	1	9300-1408
Fraction Collector Capillary Assembly (25 µm ID)	1	G1364-87304
Fraction Collector Capillary Assembly (100 µm ID)	1	G1364-87306
ZDV Union, stainless steel	1	5022-2184
Strip with 10 MALDI Spotting Tips	1	G1364-81701
Plastic Spacers	20	not orderable
Plug for waste container	1	G1364-26105

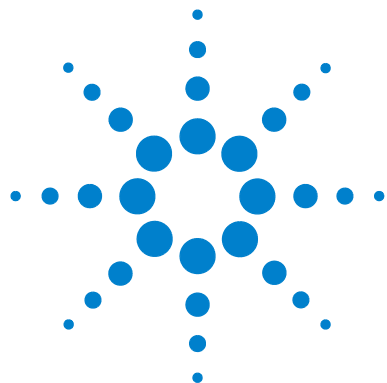
* Reorder gives 5 m

† Reorder gives 10 clamps for corrugated waste tubing (large) and 10 clamps for waste tubing (small)

‡ Reorder gives 10 clamps for corrugated waste tubing (large) and 10 clamps for waste tubing (small)

6 Parts and Materials

Micro Collector/Spotter Accessory Kit



7 Specifications

Performance Specifications for the Micro Collector/Spotter 140



Performance Specifications for the Micro Collector/Spotter

Table 20 Performance Specifications Agilent 1200 Series Micro Collector/Spotter (G1364D)

Type	Specification
trigger modes	Time slices, Peak (threshold, up- /downslope), Timetable (combination of time intervals and peak) and Agilent 1200 UV-Vis detectors DAD G1315A/B, MWD G1365 A/B are fully supported. Other detectors with appropriate delay volumes can be connected through UIB interface.
operating modes	Above location Into location Liquid Contact Control: The tip of the fraction collector capillary will initially move down to the bottom of the well. Then it will slowly move upwards while the fraction is collected. The contact control mode enables the micro collector/spotter to collect fractions down to 2 µl in well plates or MALDI spots down to 100 nl
fraction vessel capacities and trays	<ul style="list-style-type: none"> • 4 well-plates full tray (MTP) with: 384 or 96-well plates (standard and conical shape) or 4 x 27, Eppendorf tubes (0.5, 1.5, 2.0 ml), MALDI Target Plates. • 2 x well-plates std. tray (MTP) + 10 x 2 ml vials (+ 1 half tray) with: 384 or 96-well plates (standard and conical shape) or 2x 27 eppendorf tubes (0.5, 1.5, 2.0 ml),
MALDI Spotting plates (pre-configured)	<ul style="list-style-type: none"> • 96 Agilent plate for AP-MALDI • 100 Applied Biosystems, 2x96 Applied Biosystems, 192 Applied Biosystems, 400 Perseptive Biosystems • Micromass 80/96 spots • Bruker Anchor Chips 384/1536 spots
MALDI Plate Capacity	4 (3 for Bruker Anchor Chip 1536)
Minimum fraction volume	Typically 2 µl (depending on the fraction collection container)

Table 20 Performance Specifications Agilent 1200 Series Micro Collector/Spotter (G1364D) (continued)

Type	Specification
MALDI spot size	100-5000 nl (depending on the MALDI plate)
maximum spotting rate	20 spots/min (1spot/3s)
Maximum flow rate	100 µl/min
delay volumes [µl]	25 µm ID fraction collector capillary: ~0.25 50 µm ID fraction collector capillary: ~1 100 µm ID fraction collector capillary: ~5
cooling	Recommended (with additional G1330B)
maximum capacity	2 micro collector/spotter connected via 2-Position, 6-Port micro valve (G1162A)
GLP features	Early maintenance feedback (EMF), electronic records of maintenance and errors
interfaces	<ul style="list-style-type: none"> - Controller-area network (CAN). - optional; LAN or external contacts interface - RS232C, - APG-remote (for remote start / stop signals to / from other modules) - Interface to G1330A Thermostat - CAN-DC-out for operation of Agilent approved external devices like valves

NOTE

Only one type of well plate or MALDI plate can be used at a time in one tray.

7 Specifications

Performance Specifications for the Micro Collector/Spotter



A Safety Information

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Solvent Information	150
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Safety Information

The following general safety precautions must be observed during all phases of operation, service, and repair of this instrument. Failure to comply with these precautions or with specific warnings elsewhere in this manual violates safety standards of design, manufacture, and intended use of the instrument. Agilent Technologies assumes no liability for the customer's failure to comply with these requirements.

General

This is a Safety Class I instrument (provided with terminal for protective earthing) and has been manufactured and tested according to international safety standards.

WARNING

If you are using flammable solvents, remove the well-plates from the tray when you turn off the sampler. You avoid the risk of building explosive gas mixtures in the tray compartment.

WARNING

After a leak in the sampler, make sure the leak plane is cleaned and dry.

Operation

Before applying power, comply with the installation section. Additionally the following must be observed.

Do not remove instrument covers when operating. Before the instrument is switched on, all protective earth terminals, extension cords, auto-transformers, and devices connected to it must be connected to a protective earth via a ground socket. Any interruption of the protective earth grounding will cause a potential shock hazard that could result in serious personal injury. Whenever it is likely that the protection has been impaired, the instrument must be made inoperative and be secured against any intended operation.

Make sure that only fuses with the required rated current and of the specified type (normal blow, time delay, and so on) are used for replacement. The use of repaired fuses and the short-circuiting of fuseholders must be avoided.

WARNING

Any adjustment, maintenance, and repair of the opened instrument under voltage is forbidden.

WARNING

Disconnect the instrument from the line and unplug the power cord before maintenance.

Do not operate the instrument in the presence of flammable gases or fumes. Operation of any electrical instrument in such an environment constitutes a definite safety hazard.





Do not install substitute parts or make any unauthorized modification to the instrument.

Capacitors inside the instrument may still be charged, even though the instrument has been disconnected from its source of supply. Dangerous voltages, capable of causing serious personal injury, are present in this instrument. Use extreme caution when handling, testing and adjusting.

Safety Symbols

Table 21 shows safety symbols used on the instrument and in the manuals.

Table 21 Safety Symbols

Symbol	Description
	The apparatus is marked with this symbol when the user should refer to the instruction manual in order to prevent risk of harm to the operator and to protect the apparatus against damage.
	Indicates dangerous voltages.
	Indicates a protected conductor terminal.
	Eye damage may result from directly viewing the light produced by the Xenon flash lamp used in this product. Always turn the xenon flash lamp off before removing it.

WARNING

A warning alerts you to situations that could cause physical injury or damage to the equipment. Do not proceed beyond a warning until you have fully understood and met the indicated conditions.

CAUTION

A caution alerts you to situations that could cause a possible loss of data. Do not proceed beyond a caution until you have fully understood and met the indicated conditions.

Lithium Batteries Information

WARNING

Danger of explosion if battery is incorrectly replaced. Replace only with the same or equivalent type recommended by the equipment manufacturer. Lithium batteries may not be disposed-off into the domestic waste.

Transportation of discharged Lithium batteries through carriers regulated by IATA/ICAO, ADR, RID, IMDG is not allowed. Discharged Lithium batteries shall be disposed off locally according to national waste disposal regulations for batteries.

WARNING

Lithiumbatteri - Eksplosionsfare ved fejlagtig handling. Udskiftning må kun ske med batteri af samme fabrikat og type. Lever det brugte batteri tilbage til leverandøren.

WARNING

Lithiumbatteri - Eksplosionsfare. Ved udskiftning benyttes kun batteri som anbefalt af apparatfabrikanten. Brukt batteri returneres apparatleverandøren.

NOTE

Bij dit apparaat zijn batterijen geleverd. Wanneer deze leeg zijn, moet u ze niet weggooien maar inleveren als KCA.



Radio Interference

Never use cables other than the ones supplied by Agilent Technologies to ensure proper functionality and compliance with safety or EMC regulations.

Test and Measurement

If test and measurement equipment is operated with equipment unscreened cables and/or used for measurements on open set-ups, the user has to assure that under operating conditions the radio interference limits are still met within the premises.

Sound Emission

Manufacturer's Declaration

This statement is provided to comply with the requirements of the German Sound Emission Directive of 18 January 1991.

This product has a sound pressure emission (at the operator position) < 70 dB.

- Sound Pressure L_p < 70 dB (A)
- At Operator Position
- Normal Operation
- According to ISO 7779:1988/EN 27779/1991 (Type Test)

Solvent Information

Observe the following recommendations on the use of solvents.

WARNING

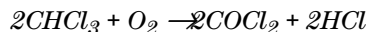
This instrument should only be used with solvents that have an ignition temperature higher than 200°C!

Solvents

Brown glass ware can avoid growth of algae.

Always filter solvents, small particles can permanently block the capillaries. Avoid the use of the following steel-corrosive solvents:

- Solutions of alkali halides and their respective acids (for example, lithium iodide, potassium chloride, and so on).
- High concentrations of inorganic acids like nitric acid, sulfuric acid especially at higher temperatures (replace, if your chromatography method allows, by phosphoric acid or phosphate buffer which are less corrosive against stainless steel).
- Halogenated solvents or mixtures which form radicals and/or acids, for example:



This reaction, in which stainless steel probably acts as a catalyst, occurs quickly with dried chloroform if the drying process removes the stabilizing alcohol.

- Chromatographic grade ethers, which can contain peroxides (for example, THF, dioxane, di-isopropylether) such ethers should be filtered through dry aluminium oxide which adsorbs the peroxides.
- Solutions of organic acids (acetic acid, formic acid, and so on) in organic solvents. For example, a 1-% solution of acetic acid in methanol will attack steel.

- Solutions containing strong complexing agents (for example, EDTA, ethylene diamine tetra-acetic acid).
- Mixtures of carbon tetrachloride with 2-propanol or THF.

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It will provide also the latest firmware of the Agilent 1200 Series modules for download.

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In This Book

This manual contains user information about the Agilent 1200 Series micro collector/spotter. The manual describes the following:

- configuration and operation,
- control module features,
- troubleshooting and test functions,
- maintenance and simple repairs,
- parts and materials,
- specifications,
- safety information.

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